Effects of breed, haemoglobin and potassium polymorphism on blood biochemical profiles of agropastoral goats

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ABSTRACT: This study was conducted to determine the effects of breed, haemoglobin (Hb) and potassium (K) types on blood profiles of agropastoral goat herds in North-Western Nigeria. A total of two hundred and fifty (250) agropastoral lactating does comprising of 109 Red Sokoto (RS), 26 Sahel (SH), 34 Kano Brown and 81 Crosses (CRS) were involved in the study. Blood was sampled from all the animals and analysed for packed cell volume (PCV), haemoglobin concentration [Hb], sodium (Na), total protein (TP), albumin (Alb), globulin (Glob) and glucose (Glu). The effects of breed, haemoglobin and potassium types on blood biochemical profile were determined using General Linear Model (GLM) procedures of SAS, significant means were separated using Duncan Multiple Range Test. The electrophoretic analysis showed three haemoglobin genotypes HbAA, HbAB and HbBB and two potassium types were observed; low potassium (LK) and high potassium (HK). The results showed that all the blood parameters measured were within the range of normal values of serum electrolytes of goats. The breeds of goats had no significant influence on blood biochemical profiles sampled except Sahel goats which had higher Total Protein (64.44±0.49 g/l) and Albumin (31.12±0.37 g/l). It was concluded that haemoglobin and potassium types had no influence on blood biochemical profile of the herd of goat studied.

Keywords: Agro-pastoral, goats, blood biochemical profile, haemoglobin, potassium.

INTRODUCTION

Goat breeds constitute the largest number of ruminants in Nigeria and are considered as perfect animals to keep due to their high ability to survive under severely harsh conditions and due to their ability to produce high-quality meat and milk (Akpa et al., 2002). The population of goats in Nigeria has been estimated to be about 53.8 million (FAOSTAT, 2008). In Nigeria, there are many breeds of goat, which include West African dwarf, Red Sokoto, Sahel, Kano brown and some crosses. Three of these breeds are most common in guinea savannah region of the country and they include Red Sokoto, Sahel, Kano brown and their crosses (Sam, 2012).

It is important to determine the blood biochemical profiles because they provide valuable information about the breed, sex and animal’s health status (Mandan et al., 2016). Several researchers have documented information on the normal values of blood parameters in domestic animal species (Ewuola et al., 2017; Isaac et al., 2013; Isidahomen et al., 2011; Alderson, 1992; Obi and Anosa, 1980; Bhergariu et al., 1984) and changes in these parameters have been studied in cattle (Ghergiri et al., 1984; Essien et al., 2011), sheep (Vihan and Rai, 1987; Gurcan et al., 2010) and goats (Tambuwai et al., 2002; Ovuru and Ekwozor, 2004; Opara et al., 2010). But the values vary according to the growth requirements, breeds and age (Piccione et al., 2007), environmental factors, managemental conditions (Arfuso et al., 2016), sexual maturity (Piccione et al., 2012), and the productivity of the animals (Mandan et al., 2016). The physiological adaptation and the systemic relationship are widely determined using the haematological values (Shah et al., 2007).

Biotechnology has opened a completely new area at molecular levels with the introduction of electrophoresis
employed for detection of polymorphism at protein and enzyme loci for the measurement of variations in animals (Salako et al., 2007). Data obtained from this type of study would be useful as genetic markers for important economic characteristics and could aid significantly in the selection of superior animals for breeding purposes (Salako et al., 2007). Haemoglobin and potassium typing are very important as different Hb and potassium types may have selective advantage in different geographical regions (Ndumukong, 1995). Blood biochemical profiles are also important and reliable for assessing the health status of individual animal (Bhat et al., 2011; Bhat et al., 2014). Despite the importance of goat in the country, very little is known on the effect of breeds on blood parameters and the relationship between hemoglobin and potassium polymorphism with blood biochemical profiles. Therefore, the aim of this study is to determine the relationship between breed, hemoglobin and potassium types with blood profiles.

MATERIALS AND METHODS

Study area

This study was carried out in Jigawa and Katsina states within the Sudan savannah zone of North-West Nigeria. Katsina lies between latitude 11°30’ to 13°01’ north of the Equator and longitude 07°41’ to 10° east of Greenwich Meridian. It is situated at an altitude of 464 m (1525 ft) above sea level. The climate varies considerably according to the season. The seasons are cool dry (harmattan) season (December to February); hot dry season (March to May); warm wet season (June to September); a less marked season after rains during the months of October to November, characterized by decreasing rainfall and a gradual lowering of temperature. The mean annual rainfall is about 780 mm. Katsina State is hot for most parts of the year, even during the wet season. Mean annual temperatures range between 19°C in the cold dry season to 38°C, with the highest temperatures normally experienced in April and May, just before the rains (Ati et al., 2007). Jigawa State falls within latitude 11° to 13°N and Longitude 8° to 10°E. The mean annual rainfall varies from 500 mm to 1000 mm. Rainfall is higher in the Southern part of the State. The mean daily maximum and minimum temperatures are 35°C and 19°C, respectively. The maximum temperature has two peaks occurring in April and October. The lowest temperatures are recorded during the month of December and January. At this period the temperature can fall as low as 10°C or lower at night. The mean relative humidity can be as high as 80% in the month of August and as low as 15% in December. The State has an altitude of between 400 to 600 m above sea level. There are two seasons in Jigawa State namely: rainy and dry seasons with the dry season lasting from November to May and the rainy season lasting between June and October (Wikipedia, 2017).

Management of experimental animals

The goats were managed under the rural agro-pastoral system of management. The animals were taken out to graze every morning from 8.00 am to 5.00 pm by children and were penned at night. On return from grazing, the animals were penned in small open sided shades by tethering. Before setting out for grazing every morning, the goats were given water and supplements which include groundnut haulms, cowpea haulms and dry grasses. Each animal was tagged with a number for individual identification. The breeds were identified by peculiar characteristics as described by Ngere (1985).

Blood collection and analysis

Five (5) ml of blood was collected from each of the sampled animals by jugular venipuncture, using sterile needle and syringe into test tube containing Ethylene Diamine Tetra Acetic acid (EDTA) anticoagulant and samples were properly labeled. Samples were carried in cold pack to haematological laboratory of Bayero University Teaching Hospital Kano, Nigeria for analysis. The blood samples were then washed with normal saline and haemolysed with distilled water to release the haemoglobin. The supernatant was removed after centrifuging at 3000 rpm for 5 minutes and the sample haemoglobin stored until ready for electrophoresis. Cellulose acetate paper strip was used to separate the globin fractions. Electrophoresis was carried out in Shandon electrophoresis tank on cellulose acetate strips 34.5 x 150 with 0.26MTris buffer (pH 9.1) at the anode and cathode. The strips were run for 5 minutes at a constant voltage of 250 V until a clear separation was observed. Interpretations were made based on the relative mobility of the haemoglobin bands towards the anode. The genotype that migrated faster was labeled HbAA, the slow moving fraction was identified as HbBB. The double band, consisting of both fast and slow band; was labeled HbAB as described by Tellah et al. (2000) and Das et al. (2004).

Haematocrit value (%) was determined by the microhematocrit method as described by Bultis and Ashwood (1994). The blood samples were centrifuged at 10,000 rpm for 5 minutes within two hours after the blood samples were collected. The haemoglobin concentration was determined by cyannmethemoglobin method. The potassium and sodium concentrations in the blood were determined by colorimetric method using spectrophotometer. The erythrocyte potassium (Ke) below or equals to 13.00 mmol/L was labeled Low Potassium (LK). Conversely, the erythrocyte potassium above 13 mmol/L was labeled High Potassium (HK) (Galip and Elmaci, 2001). Total protein was obtained using Biuret method as explained by Tietz (1995). Glucose level was determined after enzymatic oxidation in the presence of glucose oxidase as described by Barham and Trinder (1972). Measurement of albumin was based on its
Table 1. Blood Biochemical profiles of goats in Katsina and Jigawa States (Mean±se).

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>No</th>
<th>Mean±se</th>
<th>CV</th>
<th>Min</th>
<th>Max</th>
</tr>
</thead>
<tbody>
<tr>
<td>PCV (%)</td>
<td>250</td>
<td>28.48±0.28</td>
<td>15.03</td>
<td>4.00</td>
<td>42.00</td>
</tr>
<tr>
<td>Hb (g/dl)</td>
<td>250</td>
<td>9.49±0.09</td>
<td>15.58</td>
<td>4.66</td>
<td>14.00</td>
</tr>
<tr>
<td>Na (Mmol/L)</td>
<td>250</td>
<td>136.40±0.42</td>
<td>6.02</td>
<td>125.00</td>
<td>141.00</td>
</tr>
<tr>
<td>TP (g/L)</td>
<td>250</td>
<td>64.44±0.49</td>
<td>12.21</td>
<td>46.00</td>
<td>94.00</td>
</tr>
<tr>
<td>Albumin (g/L)</td>
<td>250</td>
<td>31.12±0.37</td>
<td>18.32</td>
<td>13.00</td>
<td>65.00</td>
</tr>
<tr>
<td>Globulin (g/L)</td>
<td>250</td>
<td>33.12±0.60</td>
<td>28.72</td>
<td>13.00</td>
<td>65.00</td>
</tr>
<tr>
<td>Glucose (g/L)</td>
<td>250</td>
<td>4.45±0.04</td>
<td>14.92</td>
<td>2.80</td>
<td>6.60</td>
</tr>
</tbody>
</table>

CV = Coefficient of variation, PCV = packed cell volume, Hb = haemoglobin concentration, Na = sodium, TP = Total protein.

Table 2. Blood biochemical profiles of goats according to location (Means ± se).

<table>
<thead>
<tr>
<th>Blood biochemical profiles</th>
<th>Jigawa</th>
<th>Katsina</th>
<th>LOS</th>
</tr>
</thead>
<tbody>
<tr>
<td>[Hb] (g/dl)</td>
<td>8.98±0.12b</td>
<td>10.27±0.13a</td>
<td>**</td>
</tr>
<tr>
<td>PCV (%)</td>
<td>26.91±0.33b</td>
<td>30.86±0.39a</td>
<td>**</td>
</tr>
<tr>
<td>K (mmol/L)</td>
<td>19.95±1.39</td>
<td>18.48±0.39</td>
<td>Ns</td>
</tr>
<tr>
<td>Na (mmol/L)</td>
<td>138.36±0.54</td>
<td>136.96±0.64</td>
<td>Ns</td>
</tr>
<tr>
<td>Total protein (g/L)</td>
<td>65.27±0.60</td>
<td>63.20±0.83</td>
<td>Ns</td>
</tr>
<tr>
<td>Albumin (g/L)</td>
<td>31.28±0.50</td>
<td>31.50±0.51</td>
<td>Ns</td>
</tr>
<tr>
<td>Globulin (g/L)</td>
<td>34.07±0.74</td>
<td>31.70±0.99</td>
<td>Ns</td>
</tr>
<tr>
<td>Glucose (g/L)</td>
<td>4.67±0.04</td>
<td>4.11±0.09</td>
<td>Ns</td>
</tr>
</tbody>
</table>

** = p<0.001, a, b = means within the same row with different superscripts differ significantly, n= no of observation, Ns = not significant, LOS = level of significance.

The data generated from the study were analyzed using General Linear Model (GLM) procedure of SAS (SAS, 2000). The model incorporated Breed, Hb-type and K-type as fixed factors while biochemical profiles were the dependent variables. Duncan Multiple Range Test (DMRT) procedure of SAS was used to separate the significant means. The linear model was as follows:

\[ Y_{ijln} = \mu + H_i + K_j + B_k + E_{ijk} \]

Where: \( Y_{ijln} \) = Measurement on traits, \( \mu \) = Population mean, \( H_i \) = Effect of \( i^{th} \) haemoglobin types (\( i = AA, AB, BB \)), \( K_j \) = Effect of \( j^{th} \) potassium types (\( j = Lk, Hk \)), \( B_k \) = Effect of \( k^{th} \) Breed (\( k = RS, BW, CRS, KB \)), \( E_{ijk} \) = Random error effect.

The t-test analysis was used to test the differences between the two locations.

RESULTS AND DISCUSSION

Blood profiles of the study goats

The means and coefficient of variation (CV) for concentration of blood biochemical profiles of the studied agro-pastoral goats are presented in Table 1. The average PCV, Hb, Na, total protein, albumin, globulin and glucose were 28.48%, 9.49 g/dl, 136.40 mmol/l, 64.44 g/L, 31.12 g/L, 33.12 g/L and 4.45 g/L, respectively. Measures of blood biochemical profile exhibited low to moderate variability (from 6.02 to 28.72). The lowest variability was recorded for Na concentration while the highest was globulin concentration. There were no significant (\( P>0.05 \)) difference between blood biochemical profiles of goats across locations (Table 2).

Breed of goats had non-significant (\( P>0.05 \)) effect on all the blood biochemical profiles measured except Na, total protein and albumin (Table 3). Na concentrations in Kano brown was significantly (\( P>0.05 \)) higher than Sahel, but statistically like other breeds. The Sahelian goats had the highest total protein and was significantly (\( P<0.05 \)) higher than the concentration in the crosses, but like the concentration in the Red Sokoto and Kano Brown. All the blood biochemical constituents measured were within the range of normal values of serum electrolytes of goats as

** Quantitative binding to the indicator 3,3,5,5-tetrabromo-m-cresol sulphophthalein (bromocresol green, BCG) (Grant et al., 1987). Globulin concentration was calculated from total protein and albumin, by subtracting albumin from total protein.**
Table 3: Blood biochemical profiles of goats (Mean values ±sd) in Jigawa and Katsina States according to breed.

<table>
<thead>
<tr>
<th>Breed</th>
<th>PCV (%)</th>
<th>Hb (g/dl)</th>
<th>Na (Mmol/L)</th>
<th>TP (g/L)</th>
<th>Alb (g/L)</th>
<th>Glob (g/L)</th>
<th>Glu (g/L)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Crs</td>
<td>26.87±0.51</td>
<td>8.96±0.18</td>
<td>133.67±0.77</td>
<td>64.09±0.91</td>
<td>31.71±0.67</td>
<td>32.56±1.10</td>
<td>4.72±0.08</td>
</tr>
<tr>
<td>Sh</td>
<td>26.67±2.57</td>
<td>8.89±0.89</td>
<td>125.67±387</td>
<td>73.00±4.55</td>
<td>35.67±3.34</td>
<td>37.33±5.53</td>
<td>4.80±0.39</td>
</tr>
<tr>
<td>RS</td>
<td>29.28±1.99</td>
<td>9.77±0.77</td>
<td>135.26±0.52</td>
<td>64.45±6.1</td>
<td>31.36±0.45</td>
<td>33.09±0.85</td>
<td>4.32±0.06</td>
</tr>
<tr>
<td>Kb</td>
<td>25.60±2.10</td>
<td>8.53±8.53</td>
<td>139.20±3.00</td>
<td>66.20±3.53</td>
<td>25.60±2.59</td>
<td>32.56±4.29</td>
<td>4.20±0.31</td>
</tr>
</tbody>
</table>

*P < 0.05; a, b, c = column means under the same factor with different superscripts differ significantly, Crs = crosses, Sh = Sahel, RS = Red Sokoto, Kb = Kano brown, PCV = packed cell volume, Hb = haemoglobin concentration, Na= Sodium, TP=Total protein, Alb = Albumin, Glob = globulin, Glu = glucose.

reported by Boyd (1984) and Radostits et al. (200).

The percentage of PCV (28.47%) in this study was like the 29.4% obtained by Daramola et al. (2005) in WAD goats. However, this is higher than 25.7% obtained in Red Sokoto goats by Tambuwal et al. (2002). Earlier reports in Baladi goats (Azab and Abdel-Maksoud, 1999) showed PCV values of 27.25%. The Hb concentration in this study (9.49 g/dl) was within the range of normal values obtained in Red Sokoto goats which is 7 to 15 g/dl (Tambuwal et al., 2002; Daramola et al., 2005). This is an advantage in terms of oxygen carrying capacity of the blood, which may lead to better functioning and performance of these animals.

The Na concentration obtained in this study (136.40 mmol/L) is within the range reported in WAD goats and Red Sokoto goats by Daramola et al. (2005) and Tambuwal et al. (2002) respectively. Sodium makes up about 93% of the basic mineral elements in the blood serum and is chief cation regulating blood pH (Swartz, 2010). The ability of muscles to contract is dependent on proper sodium concentration. Therefore, these indigenous goats could be said to have normal sodium concentration in the blood. Total protein in this study (64.44 g/l) was within the range reported by Tambuwal et al. (2002) in Red Sokoto goats and Oduye and Adadevoh (1976) reported same in WAD sheep. Lower concentration of albumin was observed in KB when compared to Sahel goat, although it was similar to other breeds sampled. Albumin concentration in this study (33.12 g/L) was within the range reported by other authors (25 to 34 g/l) (Oduye and Adadevoh, 1976; Tambuwal et al., 2002; Daramola et al., 2005).

Proteins are the basic components of enzymes, many hormones, antibodies and clotting agents. They are substance that aid in the transportation of hormones, vitamins, minerals, lipids and other materials (Kaslow, 2011; Njidda et al., 2014). In addition, proteins help balance the osmotic pressure of the blood and tissue (Kaslow, 2011). Total protein in the blood which represents the sum of albumin and globulin is a very strong indicator of the health status of the animals; low albumin is a sign of poor health (Ewuola et al., 2017). Serum total protein is not related to the amount of calories contained in diets but to the availability of protein for utilization (Tewe and Maner, 1980). Globulin and glucose concentration were similar for all the breeds sampled. The glucose concentration (4.44 g/l) observed in this study is higher than 2.3 mmol/L and 3.9 mmol/L in the study of Jovanovic et al. (1998) and McDongall et al. (1991), respectively. Zubic (2001) established that glucose level decreases with age of goats, and most of the animals in this study were between ages 2 to 6 years which probably explains the high concentration of glucose. Glucose is the major precursor of lactose synthesis in the mammary gland, and lactose is the major carbohydrate and osmolyte of milk which controls milk volume and its concentration (Shahbazkial et al., 2010).

Effect of haemoglobin and potassium polymorphism on blood profiles in goats

Table 4 shows blood profiles of goats based on haemoglobin and potassium types. The Hb type significantly (P<0.05) influenced PCV; animals with HbAA had significantly (P<0.05) higher PCV values than those with HbAB and HbBB which were statistically similar. Haemoglobin concentration (Hb) was also significantly influenced by Hb types, animals with HbAA had significantly (P<0.05) higher concentration of haemoglobin (Hb) than animals with HbAB and HbBB. Hb-type had no significant (P>0.05) influence on Na, total protein, albumin, globulin and glucose. On the other hand, potassium types have no significant (P>0.05) effect on all the blood profiles measured except on Na. HK animals had significantly (P<0.05) lower Na concentration than LK potassium animals.

In the present study, Hb type only influenced packed cell volume (PCV) and haemoglobin concentration, animals with HbAA recorded higher values of PCV than animals with HbAB and HbBB. Carriers of haemoglobin AA genotype have been reported to have better functional properties such as affinity for oxygen, higher haemoglobin concentration and packed cell volume (FAO, 2008; Akinnyemi and Salako, 2010).

The effect of potassium types significantly influenced Na concentration. It had been observed that HK animals had less concentration of Na than LK (Galip and Elmaci, 2001). This observation agrees with the reports of Gurcan et al.
(2010) that noted significant difference in the mean Na concentration of HK and LK, which they observed to be 174 and 150 mmol/l for LK and HK, respectively. This variation could be attributed to Na – K pump sites per unit cell membrane surface. Shahbabak et al. (2009) explained that HK type animals had low number of Na – K pump sites which lead to a lower concentration of Na in the cell while HK type animals have a higher number of Na – K pump sites, hence the increase in Na concentration of these animals.

**Conclusion**

The blood profiles of the studied Agro-pastoral goats were within the reference normal range for goats. The locations of animals have no influence on all the blood parameters analyzed, except on Hb concentration and PCV. The breeds of goats have no significant influence on blood profiles sampled, except in the Sahel goats which had higher total protein and albumin; but lowest Na compared to other breeds. Haemoglobin and potassium types also have no influence on all the blood parameters analysed.

**CONFLICT OF INTEREST**

The authors declare that they have no conflict of interest.

**REFERENCES**


