

Insects associated with green gram (*Vigna radiata* L.) in Langtang, Southern Guinea Savanna of Nigeria

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ABSTRACT: Green gram is a 'wonder' or 'super' food with health benefits. It is one of the shortest growth duration grain legumes that are also suitable as catch crops. Leguminous crops generally attract insect pests because of their high nutritive value. Hence, a survey of insects associated with green gram (*Vigna radiata* L.) treated with insecticide and fungicide was conducted in Kwanpe, a suburb of Langtang in Plateau State, Nigeria from July to September during the 2014 cropping season. Four treatments of dimethoate (an insecticide) were applied (at 0, 300, 500, and 700 g a.i./ha) in four replicates arranged in a Randomized Complete Block Design. A total of 40 insect species belonging to 4 orders and 8 families were captured. Species known to be pests in other legume crops encountered were 22. There were few predatory species. There were no statistical differences in pest species populations in treated and untreated green gram plots. It is recommended that there is no need to engage synthetic chemicals in the production of green gram in this locality. Regular insect surveillance is however required as a monitor for pest species populations in order to set economic threshold levels in this Southern Guinea Savanna ecological zone for the highly valued crop.

Keywords: *Vigna radiata*, crop damage, insect checklist, pest control.

INTRODUCTION

Majority of people in the developing countries are engaged in agricultural activities but with low productivity (Kumar, 1991). Grain legumes are of considerable importance in Nigeria because they provide a major source of protein in human and animal nutrition. They are of particular advantage in subsistence and mixed farming because they can grow on nitrogen-deficient soils (Schroth et al., 2000; Rao et al. 2004). They are well fitted to agro-forestry cropping systems (Rao et al., 2000). Green gram grows best at an altitude of 0 to 1600 m above sea level and under warm climatic conditions (temperature range of 28 to 30°C). They are well adapted to soils that are red sandy loam, but also do reasonably well on those that are not too exhausted though sandy. Green grams are not tolerant to wet, poorly drained soils. They are drought tolerant and will give reasonable yields with as little as 650 mm of yearly rainfall. Heavy rainfall results in increased vegetative

growth with reduced pod setting and development (CBS Kenya Govt, 2003; URT, 2003). Additionally, it is adapted to poor soils because it forms associations with mycorrhiza (Kasiamdari et al., 2002) and is a relay crop, hence plays an important role in environmental conservation and food security, respectively (Machocho et al., 2012).

However, production of green gram is constrained by diseases, pest infestations, unsuitable varieties and inappropriate agronomic practices (Rao et al., 2000). These practices, particularly pest and disease control, entail the use of industrial chemicals. The practices are expensive, pose health hazards and are environmentally undesirable. Besides, the chemicals are not popular among the resource poor farmers who are also the main producers and consumers of the legume (Machocho et al., 2012).

The major constraint to current legume yields in India and some parts of East Africa is pest infestation, either as vectors of diseases or destroyers of seedlings, foliage or fruiting bodies in India (Davies and Lateef, 1975; Saxena, 1978; Seif et al., 2001). Incidences of diseases and pests are comparatively low during the dry months when the crop escapes rain damage. According to Davies and Lateef (1975), for instance, pests account for 20 to 25% loss of green gram yield in India. It has been established that in order to achieve high crop yields, even with the potentially high yielding varieties grown in the United States of America, plants must be adequately protected from insect pests (Cook, 1986).

Green gram is one of the shortest growth duration grain legumes that are also suitable as catch crops (Seif et al., 2001; Emmanuel et al., 2017). It arrests the hunger which peasant farmers often experience, where it is grown, before many other crops mature (Purseglove, 2003; Anon, 2007). While the world production of *V. radiata* is concentrated in India and China, records of production on the African continent are in Kenya and Tanzania. In Nigeria, the crop is highly restricted to Langtang-North and Wase Local Government Areas of Plateau State and said to have been introduced by the white missionaries of the Sudan United Mission (SUM) in 1908/1909 (Lere, 1996). The crop was a readily available source of vegetable eaten with boiled rice. For over a centenary now, farmers in the locality have not stopped producing and consuming the crop on small scale. Production is however gradually gaining commercial quantity (Seif et al., 2001).

The crop is referred to as a 'wonder' or 'super' food (Mogotsi, 2006). The health benefits of the beans are increasingly exploding with more research on the crop. Weight loss is achieved because the beans are low in calories and rich in fibre (Minh, 2014). The insoluble fibres present in the seeds help to keep the digestive system healthy and reduce the problem of constipation. Eating a small cup of green gram bean soup therefore gives the feeling of fullness. It therefore helps to curb hunger pangs and bring body weight to healthy levels. Regular consumption of the seeds helps to reduce unhealthy cravings for sweet artificial foods thus regulate blood sugar level (Arya, 2014). Its high potassium content lowers blood pressure by counteracting the effect of sodium (Mogotsi, 2006; Arya, 2014).

It has an awesome and acceptable taste no matter the preparation. In India, the bean is cooked for sick people who may have lost appetite for food. Its healing properties have therefore been found to be gaining undisputable popularity. The potassium and magnesium components of the seeds are important for a healthy heart, while its folic acid is important for pregnant women and women of child bearing age (Arya, 2014). Other minerals present in the crop include zinc, iron and phosphorus. Vitamins are essential to maintain good health and prevent diseases. Green mung bean sprouts are rich in Vitamin C which improves the immune system and keeps the common

fever, sore throat and cold away. Eating green gram thus gives the benefit of fruit consumption (Minh, 2014; Raman, 2018; Umata, 2018).

A host of benefits of consuming green gram was outlined by beneficiaries as: it helps to reduce weight and fight obesity, lowers blood pressure, controls cholesterol and heart disease risk, helps fight cancer, boosts immunity and protects against infections, improves skin health and possesses anti-toxic properties (Arya, 2014). In China and India, green gram beans are frequently recommended to detoxify the blood and get rid of chronic illnesses. It is an anti-inflammatory food and helps to heal the body through improved body metabolism (Tang et al., 2014).

Leguminous crops generally attract insect pests because of their high nutritive value (Mogotsi, 2006). Their protein and vitamin content are a source of nourishment to insects, just as humans. In Uganda both yield and seed quality are significantly reduced (ranging from 80-100%) by damage due to insects, if no control is undertaken (Emmanuel et al., 2017). Adamu et al. (2001) recorded up to 92% seed losses in green gram due to damage by the pod weevil, *Piezotrachelus varius* in the northern guinea savanna zone of Nigeria. Whiteflies (*Bemisia tabaci* and *Aleurodicus dispersus*), blister beetles (*Mylabris species*) and stink bug (*Nezara viridula*) were also found in the area.

Since green gram have economic, nutritive and medicinal values, however there is paucity of information on insect fauna of the plant. Hence, the aim of the study was to survey insects that are associated with the crop in its presently well adopted area of production in relation to treatments with dimethoate insecticide and fungicide. It is hoped that the findings in this experiment would provide useful information to further study the insect fauna of the crop as its production is fast increasing.

MATERIALS AND METHODS

The study was carried out in Kwanpe, a suburb of Langtang (latitude 9°08'N: longitude 9°47'E) in Plateau State, Nigeria from July to September during the 2014 cropping season. A total land area of 320 m² (20 m x 16 m) was ploughed and harrowed. Compound fertilizer (N₁₅ P₁₅ K₁₅), was applied at the rate of 200mkg/ha, according to the common practice of green gram farmers, before ridges of 100 cm apart were made. Four blocks separated by 1 m alleys and consisting of four plots (4 cm x 3 cm) which were each separated by a 1 m alley to minimize inter-plot interference were provided.

Four treatments, consisting of three levels of insecticide application and a control, were replicated four times. These were arranged in a Randomized Complete Block Design (RCBD). Dimethoate, an organophosphate insecticide available as Kartodim 315 EC, was applied at the rate of (0, 300g, 500 and 700 g a.i./ha). A broad spectrum preventive fungicide for the control of foliar fungal diseases, copper oxychloride, available as Red

Force was tank-mixed at the rate of 800 g a.i./ha with dimethoate and sprayed using a knapsack sprayer. Planting was done in the last week of July 2014. Pesticides application commenced two weeks after planting (WAP) (i.e. 2nd week of August 2014) and continued at weekly intervals thereafter till week 8. Control plots received water spray only. Spraying time was maintained at 9.00 hours each time.

Sampling of Insect fauna

The Sweep Net: Insects were collected from 9.00 to 11.00 hours using a sweep net of 18 cm diameter, 28 cm deep, mesh size of about 20 mm and a handle 29 cm long. Five swings along 3 m-row of crop canopy on one of two middle rows in each plot constituted one sweep, according to the method of Paulson (2005). After each sweep-net sampling, net was emptied into a killing jar. Insect specimens were immediately transferred to labeled transparent specimen bottles containing 70% alcohol for preservation. They were conveyed to the laboratory for identification.

Tagged Plants: Five plants per plot were randomly tagged for weekly *in situ* observation. Tagged plants were closely observed for sedentary insects same day with sweep net sampling. With the aid of flexible forceps any insect observed was picked, killed and preserved in 70% alcohol. Collections were conveyed to the laboratory for identification.

Assessment of growth and yield components of the crop

Crop growth and yield parameters assessed were; number of leaves per plant per week, number of peduncles per plant, number of days to first flower, number of days to 50% flowering, number of pods per plant, number of insect-damaged seeds per plant, 100 seed weight and grain yield (Dike, 1997).

Insect-damaged seeds were recorded by observing insect exit holes on seeds or scarified seed coat or when the germ of the seed was eaten by insects. One hundred seeds picked randomly from the harvested pods of two middle rows of each plot were weighed. Grain yield was measured by bulking pods from two middle rows of each plot, threshing and then weighing them (Dike, 1997).

Insect identification

Insects collected by both sweep net and hand-picking from tagged plants were identified to species level using morphological characteristics by comparing with preserved specimens at the Insect Museum of the Ahmadu Bello University, Samaru, Zaria. Collections at the Insect Museum had been carefully identified using standard keys which include Borror and White (1970), Skaife (1979), Castner (2000) and Shattuck (2000).

Statistical analysis

Data obtained was analyzed using R-Console Software version 3.2.2. Pearson's Chi-squared test was used to compare the abundance of insects in relation to treatments on *Vigna radiata*. Also, the proportion of number of days to flowering, number of days to 50% flowering, number of pods per plant, weight of 100 seeds, number of insect-damaged seeds and as well as the mean grain yield in relation to treatments on *Vigna radiata* were compared using Chi-square. P-values < 0.05 were considered statistically significant.

RESULTS

Composition of insects associated with green gram

Tables 1 and 2 show the number of insect species captured in this experiment, listed according to their perceived roles on the crop. Only pest species are shown in Table 1 where 5 insect orders were present. Hemiptera and Coleoptera had 11 and 8 species, respectively. Two species of Orthoptera, one of Homoptera and one of Diptera were recorded.

Table 2 shows the insect orders with predatory and parasitoid species. They included Hymenoptera (4 species), Coleoptera (5 species), Dictyoptera (2 species), Heteroptera (2 species each) and Diptera (2 species). Two species; *Lebialate plagiata* (Coleoptera: Carabidae) and *Cardiophorus hoploderus* (Coleoptera: Elateridae) could not be grouped with certainty.

Table 3 shows the overall relative abundance of species. The most abundant were *Mirperus jaculus* and *Monolepta nigeriae*, constituting 13.71 and 13.19% of the total insect population captured, respectively. *Estcourtiana bifasciata* and *Chilomenes sulphurea* closely followed in abundance at 12.09 and 9.34%, respectively.

The number of insects caught was highest (11) in control followed by 300g and 500g dimethoate treatments in which both had 10 while the 700g had the least 9 as shown in Table 4. However, there was no significant difference ($\chi^2 = 0.2$, df = 3, P = 0.9776) in the number of insects caught among the treatments.

Table 5 shows the abundance of insect orders in descending sequence. The most abundant groups were Coleoptera and Hemiptera. Relative abundance of insects according to families is shown in Table 6. Chrysomelid beetles constituted 35.6% of the total captured insects.

Plant growth and yield parameters

Figure 1 shows the number of leaves per plant for 8 weeks. The maximum and minimum leaf counts were 51 and 47 for the control and dimethoate at 500 g a.i./ha plots, respectively. These indicated there were no appreciable differences ($\chi^2 = 0.20408$, df = 3, P = 0.9769) in leaf

Table 1. Insect pests associated with *Vigna radiata* Langtang, Plateau State, Nigeria

Order/Name of Insect	Family
Coleoptera	
<i>Monolepta nigeriae</i> Bryant.	Chrysomelidae
<i>Monolepta gossypiperda</i> Bryant.	Chrysomelidae
<i>Monolepta kraatzi</i> Jac	Chrysomelidae
<i>Estcourtiana bifasciata</i> Jac	Chrysomelidae
<i>Cheilomenes sulphurea</i>	Coccinellidae
<i>Lema tibialis</i> Casteln.	Chrysomelidae
<i>Aulacophora africana</i> Weise.	Chrysomelidae
<i>Nisotra dilectra</i> Dalm.	Chrysomelidae
Hemiptera	
<i>Lygaeus rivularis</i> Germ.	Lygaeidae
<i>Aspavia acumianta</i> Mont.	Alydidae
<i>Nezara viridula</i> F.	Pentatomidae
<i>Aethemenes chloris</i> Westw.	Pentatomidae
<i>Poophilus costalis</i> Walker.	Pentatomidae
<i>Clavigrallato mentosiculis</i> Stal	Cercopidae
<i>Anoplocnemis curvipes</i> Fabr.	Coreidae
<i>Agonoscelis versicolor</i> F.	Pentatomidae
<i>Mirperus torridus</i> Westw.	Alydidae
<i>Halydicoris species</i>	Pentatomidae
<i>Phorticus flavus</i> Stal.	Nabidae
<i>Locris rubens</i> Erichson	Cercopidae
Orthoptera	
<i>Tylopsis irregularis</i> Karsch.	Tettigoniidae
<i>Melanostictus melanostictus</i> Schaun.	Acrididae
Diptera	
<i>Tricyclea species</i> Casteln.	Calliphoridae

production due to insecticide application.

It took between 35 and 39 days for the first flower to appear in all the treatments. There was no significant difference ($\chi^2 = 0.2349$, df = 3, P = 0.9718) in the number of days for the first flower to appear between treatments (Table 7). Similarly, there was no significant difference ($\chi^2 = 0.47191$, df = 3, P = 0.925) between treatments in the number of days (41 to 47 days) it took to obtain 50% flowering (Table 8).

Figure 2 is a record of the number of peduncles per plant over time. Up to 15 peduncles per plant was recorded and in plots treated with dimethoate at 500 g a.i./ha while the least number of peduncles (average of 13) was in plots treated with insecticide at 300 g a.i./ha. However, the number of peduncles among treatments in relation to each observed week showed no significant difference (Week 4: $\chi^2 = 0.4$, df = 3, P = 0.9402; Week 5: $\chi^2 = 0.14286$, df = 3, P = 0.9862; Week 6: $\chi^2 = 0.28205$, df = 3, P = 0.9634; Week 7: $\chi^2 = 0.14286$, df = 3, P = 0.9862).

The number of pods per plant (Table 9) ranged from 72 to 79, the lowest being in the control plot and the highest was in dimethoate plot treated with 500 g a.i./ha. However, the number of pods per plant across treatments showed no significant difference ($\chi^2 = 0.39474$, df = 3, P = 0.9413).

Effect of insect pest damage on yield of *Vigna radiata*

Figure 3 compared the number of insect-damaged and undamaged seeds between treated and untreated plots of *V. radiata*. The number of damaged seeds in all cases was negligible. There was no significant difference in the number of insect-damaged seeds between treatments on *V. radiata* ($\chi^2 = 1.0886$, df = 3, P = 0.7819).

Each of the treatments had same weight (5 g) for 100 seeds. Grain yield ranged between 0.0015 and 0.0017 t/ha (Table 10) and were not significantly different ($\chi^2 = 0.00001746$, df = 3, P = 1) across treatments.

Table 2. Predators and parasitoid associated with *Vigna radiata* in Langtang.

Feeding Guild	Name of Insect	Family
	Coleoptera	
Predator	<i>Paederus sebaeus</i> Er.	Staphylinidae
Parasitoid	<i>Lebia malanura</i> Dej	Carabidae
Predator	<i>Stenidia corrusca</i> Lar.	Carabidae
Predator	<i>Pachnoda cordata</i> Dry.	Scarabaeidae
	<i>Diplognatha gagates</i> Forst.	Scarabaeidae
	Dictyoptera	
Predator	<i>Mantis</i> species	Mantidae
	<i>Sphodromantis</i> species	Mantidae
	Hemiptera	
Predator	<i>Rhinocoris segmentarius</i> Germ.	Reduviidae
	<i>Rhinocoris tropicus</i> H&S	Reduviidae
	Hymenoptera	
	<i>Camponotus sericeus</i> Fab.	Formicidae
Predator	<i>Pheidole</i> species	Formicidae
	<i>Braunsia analis</i> Kriechb.	Braconidae
	<i>Camponotus acvapimensis</i> Mayr.	Formicidae
	Diptera	
Predator	<i>Morellia</i> species	Muscidae
	<i>Musca domestica</i> (L.)	Muscidae

Table 3. Cumulative relative abundance of insects on *Vigna radiata* (L.).

Insect species	Number	Relative abundance (%)
<i>Mirperus jaculus</i>	25	13.74
<i>Monolepta nigeriae</i>	23	13.19
<i>Estcourtiana bifasciata</i>	22	12.09
<i>Cheilomenes sulphurea</i>	17	9.34
<i>Nezara viridula</i>	8	4.40
<i>Monolepta kraatzi</i>	8	4.40
<i>Lebia melanura</i>	7	3.85
<i>Pheidole</i> species	6	3.30
<i>Anoplectnemis curvipes</i>	5	2.75
<i>Aulacophora africana</i>	5	2.75
<i>Agonoscelis versicolor</i>	4	2.10
<i>Lygaeus rivularis</i>	3	1.65
<i>Mirperus torridus</i>	3	1.65
<i>Camponotus sericeus</i>	3	1.65
<i>Musca domestica</i>	3	1.65
<i>Diplognatha gagates</i>	3	1.65
<i>Paederus sebaeus</i>	2	1.10
<i>Monolepta gagates</i>	2	1.10
<i>Phorticus flavus</i>	2	1.10
<i>Aethemenes chloris</i>	2	1.10
<i>Cardiophorus hoploderus</i>	1	0.55
<i>Sphodromantis</i> sp.	1	0.55

Table 3. Contd.

<i>Mantis</i> sp.	1	0.55
<i>Tricycloa</i> sp.	1	0.55
<i>Lema tibialis</i>	1	0.55
<i>Camponotus acvapimensis</i>	1	0.55
<i>Halydicoris</i> species	1	0.55
<i>Clavigralla tomentosicollis</i>	1	0.55
<i>Lebia lateplagiata</i>	1	0.55
<i>Melanastictus melanastictus</i>	1	0.55
<i>Morellia</i> species	1	0.55
<i>Nisotra dilecta</i>	1	0.55
<i>Stenidia corrusca</i>	1	0.55
<i>Tylopsis irregularis</i>	1	0.55
<i>Rhinocaris tropicus</i>	1	0.55
<i>Aspavia acuminata</i>	1	0.55
<i>Poophilus costalis</i>	1	0.55
<i>Locris rubens</i>	1	0.55
<i>Agonocelis</i> species	1	0.55
<i>Pachnoda cordata</i>	1	0.55
<i>Braunsia analis</i>	1	0.55
<i>Rhinocoris segmentarius</i>	1	0.55
Total	174	100

Table 4. Total number of insects per treatment in the experiment

Dimethoate (g a.i./ha)	Number of insects
Control	11
300	10
500	10
700	9

$\chi^2 = 0.2$, df = 3, P = 0.9776.

Table 5. Total abundance of insect orders associated with *Vigna radiata* L. in Langtang during the 2014 cropping season.

Insect order	No. of insects	Percentage
Coleoptera	94	54.0
Hemiptera	61	35.0
Hymenoptera	10	5.7
Diptera	5	2.9
Dictyoptera	2	1.2
Orthoptera	2	1.2
Total	174	100

DISCUSSION

Insect composition and association with green gram

In all, 40-insect species were identified associated with the crop. This association may have evolved over time as no

Table 6. Relative abundance of insect families associated with *Vigna radiata* during the study period.

Family	Percentage
Chrysomelidae	35.6
Alydidae	16
Coccinellidae	9.8
Pentatomidae	9.8
Formicidae	5.8
Carabidae	5.3
Coreidae	3.4
Scarabaeidae	2.3
Muscidae	2.3
Lygaeidae	1.7
Staphylinidae	1.3
Mantidae	1.3
Nabidae	1.3
Cercopidae	1.3
Elateridae	0.5
Calliphoridae	0.5
Acrididae	0.5
Tettigoniidae	0.5
Reduviidae	0.5
Braconidae	0.5
Total	100

entomological findings had been carried out on the crop in this ecology since the introduction of the crop over a century ago. The dominance of *Mirperus jaculus* and

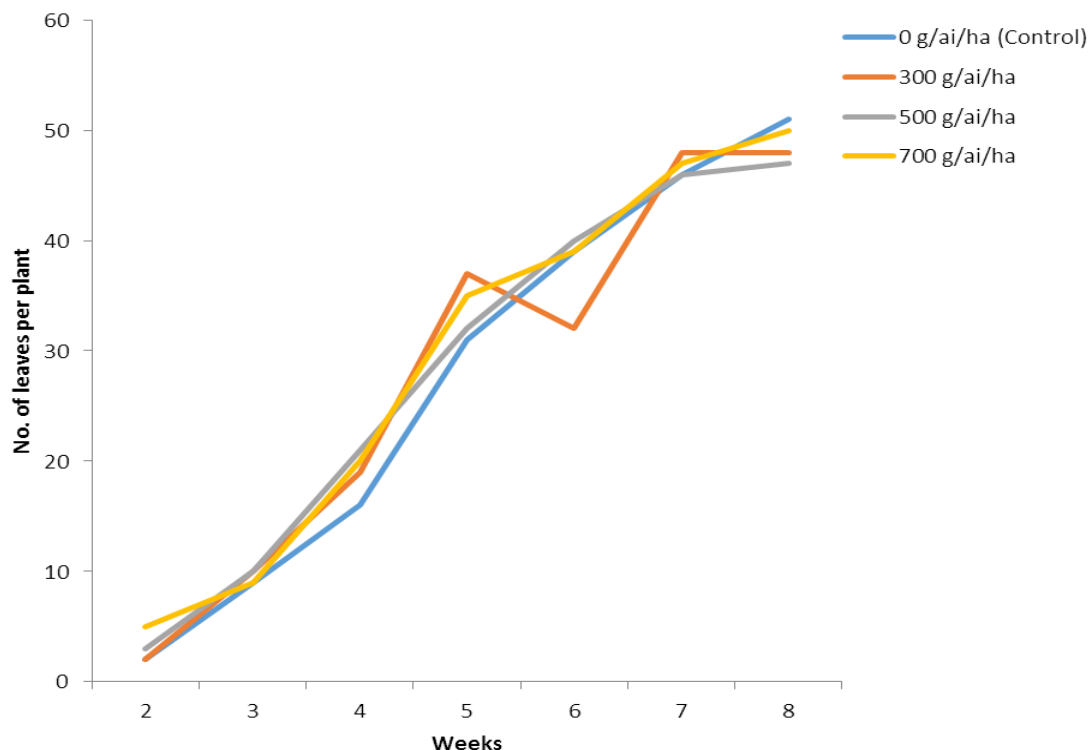


Figure 1. The number of leaves per plant on insecticide treated *Vigna radiata* plants.

Table 7. Number of days to first flower emergence of insecticide treated green gram plants.

Dimethoate spray (g a.i./ha)	Number of flowering day
Control	35
300	37
500	39
700	38

$\chi^2 = 0.2349$, df = 3, P = 0.9718.

Table 8. Number of days to 50% flowering of insecticide treated green gram.

Dimethoate spray (g a.i./ha)	Number of days to 50% flowering
Control	41
300	44
500	47
700	46

$\chi^2 = 0.47191$, df = 3, P = 0.925.

Monolepta nigeriae over other species suggests that there are potential pests of the crop in this sub-northern guinea savanna ecological zone. This corroborates the findings of Ndam (2012) on *Mirperus jaculus* and *Monolepta nigeriae*

which were found to have associated with soybean (another legume crop) for many years in Benue State within the same ecological zone. Also, Seif et al. (2001) showed that French beans are attacked by a number of different pests which include bean flies, bean flower thrips, aphids, pod borers, bean rust, red spider mites. The severity of infestation of different pests varies depending on the location and season.

Insect abundance

The lack of variation in the number of insects among the treatments possibly suggests that there is some form of insecticide resistance in the insects in relation to location and season. This is in accordance with the finding of Seif et al. (2001) who showed that the severity of infestation of different pests varies depending on the location and season as farmers control various pests by application of foliar pesticides, especially insecticides in Kenya.

The abundance of insects among 0 g (control), 300, 500 and 700 g a.i./h treatments in descending order was 11, 10, 10 and 9 respectively. This suggest that plot with the highest concentration of insecticide (700 g a.i./h) tend to repel and influence low insects abundance on *V. radiata*. This is accordance with the finding of Stewart and McClure (2018) who showed that insecticide resistance by insects can be attributed to level of insecticide dosage. Therefore,

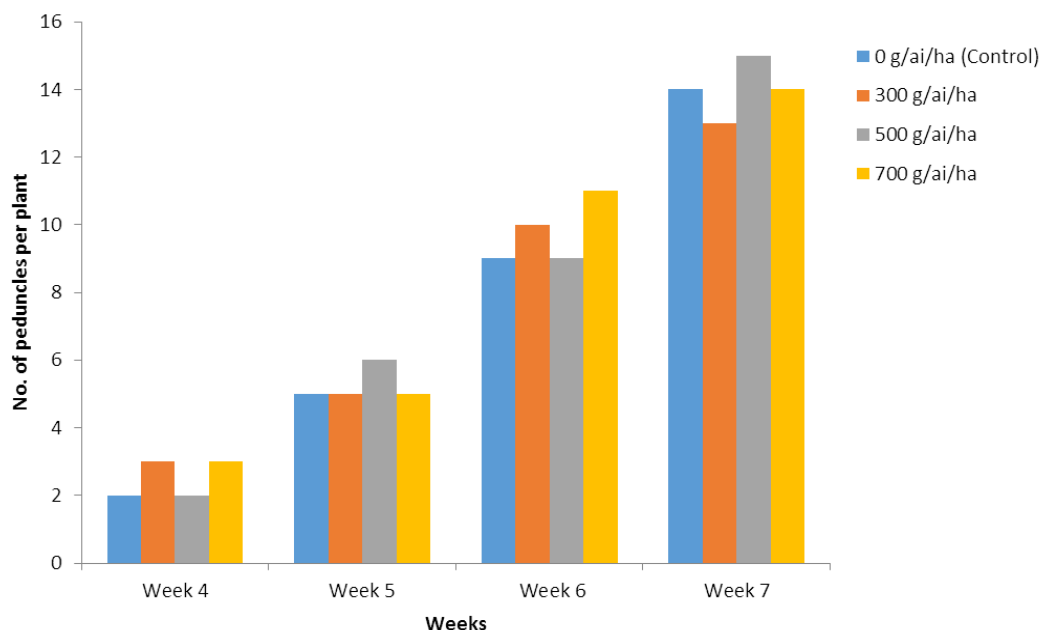


Figure 2. Number of peduncles per plant on insecticide treated *Green Gram*.

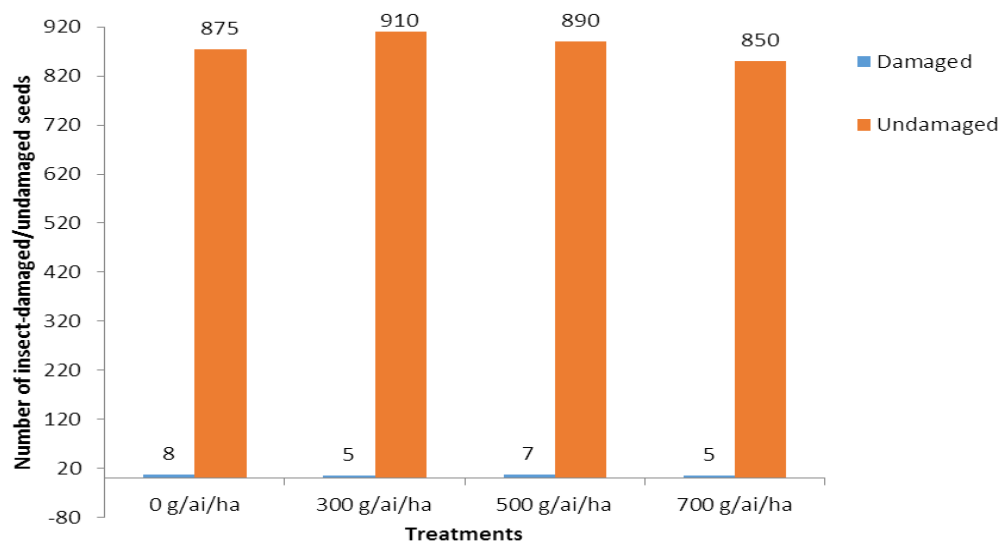


Figure 3. Number of insect-damaged and undamaged seeds from insecticide-treated green gram plants.

increase in insecticide dosage will take care of unsatisfactory control problems of foliar insects.

The total insect count on the crop at the end of the experiment is low. Coleopterous and hemipterous pest species are most abundant. The fact that 54% of all the insects caught belong to the order coleoptera, gives high hope for natural control. Singh and Taylor (1978) emphasized that predators, especially coccinellid beetles, keep the population of *Aphis craccivora* under control in the derived savanna ecology in south western Nigeria.

Effect of insecticide application on plant growth parameters

The number of days to first flower emergence (35 to 39) is not affected by insect infestation as both treated and untreated plots develop at the same place. Similarly, all other growth and yield parameters; number of leaves per plant, number of days to 50% flowering, number of peduncles per plant and number of pods per plant indicate that green gram could be produced without an insecticide application.

Table 9. Number of pods per plant on insecticide treated green gram.

Dimethoate spray (g a.i./ha)	Number of pods per plant
Control	72
300	75
500	79
700	78

$\chi^2 = 0.39474$, df = 3, P = 0.9413.

Table 10. Grain yield of insecticide treated green gram.

Dimethoate spray (g a.i./ha)	Grain yield (t/ha)
Control	0.0017
300	0.0015
500	0.0015
700	0.0016

$\chi^2 = 0$, df = 3, P = 1.

Effect of insect damage on yield

All yield parameters; number of insect-damaged seeds, 100-seed weight and grain yield are indicative of minimal negative impact of pest species on the crop. While pesticides are meant to provide good protection against pod infestation by pod borers for better seed yield (Mugo, 1989; Muthomi et al., 2007), findings in this experiment are not corroborative.

Some environmental factors that are beyond the control of the researchers may have accounted for lower yields in the dimethoate-treated plots than control, as also observed by Okeyo-Owuor (1978).

Conclusion

The production of green gram in the Langtang location is not yet threatened by the insect pest species identified and collected. The crop is however exposed to the danger of pest attack as environmental dynamics take their toll. The research further affirms that the area is very suitable for the production of the crop.

Recommendation

It is therefore recommended that insect surveillance be sustained to monitor insects associated with green gram in this area. This would ameliorate cases of pest emergence among potential pests, unnoticed. It would further help identify possible changes in pests that may be occasioned by changes in consumers' demand for the crop. This recommendation is in line with the emphasis PROTA (2017) has placed on the crop for further research to improve on its productivity.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

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