Effect of cowpea enrichment on the physico-chemical, mineral and microbiological properties of maize:cowpea flour blends

Shakpo, I. O.1,2* and Osundahunsi, O. F.2

1Food Science and Technology Department, Rufus Giwa Polytechnic, Owo, Nigeria.
2Food Science and Technology Department, Federal University of Technology, Akure, Nigeria.

*Corresponding author. Email: de.kings.io@gmail.com. Tel: +2348036267170.

Copyright © 2016 Shakpo and Osundahunsi. This article remains permanently open access under the terms of the Creative Commons Attribution License 4.0, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Received 25th February, 2016; Accepted 20th June, 2016

ABSTRACT: Production of nutritious low cost complementary foods is a panacea to malnutrition in developing countries. The objective of this study was to examine the complementary effect of cowpea on some quality attributes of maize flour. Flour blends were produced from maize and cowpea flours in the following ratios of maize: cowpea; 90:10, 80:20, 70:30 and 100% maize as control. Physicochemical (proximate, mineral and functional properties) and microbiological analyses were carried out on the flour blends. The proximate result showed that the protein content ranged from 8.85 to 10.52%, total ash, 1.55 to 1.93%; fat content, 10.50 to 11.96%; moisture content, 5.05 to 5.77% and fibre content, 9.88 to 14.43%. The blend with 30% cowpea substitution gave the highest value. It was observed that fat content was not affected by cowpea substitution. Mineral determination showed that potassium, sodium, zinc, and iron contents increased with cowpea substitution in the blends, with 30% cowpea substitution having the highest values. Calcium contents were 3.34, 3.36, 3.39 mg/kg for 10%, 20% and 30% cowpea substitution respectively; while calcium was not detected in 100% maize. Functional properties evaluated showed that bulk density ranged from 0.72 to 0.81 g/ml, swelling index, 0.66 to 1.03 ml/g, least gelation, 2.10 to 6.11%, foaming, 7.67 to 15.36%, water absorption, 2.41 to 3.23%, oil absorption 1.24 to 1.45%, emulsion capacity, 8.76 to 37.15% and emulsion stability 7.43 to 14.13%. Microbiological evaluation for the flour blends showed that total viable count ranged from 3.80 to 4.7 x 10^3 CFU/ml, while yeast and mould count ranged from 2.2 to 3.17 x 10^3 CFU/ml for blends. The overall result showed that 20% cowpea substitution is the most adequate percentage to produce an acceptable and nutritious flour blend from maize and cowpea which can be useful for pastries and confectioneries.

Keywords: Cowpea, enrichment, maize, malnutrition, physico-chemical.

INTRODUCTION

The world cultivated cereals include wheat, maize, rice, barley, Oat, rye, sorghum, millet etc., an important characteristic of cereals is that they have high carbohydrate, low fat and a fair content of protein (Olugbani et al., 2002). Wheat is a popular cereal grain that has been used for the production of many snacks, such as chin-chin, biscuit, pie, bread and other pastry products (David, 2006) but the tropical climate of many developing countries does not encourage commercial wheat cultivation, leading to reliance on wheat importation. Recently in Nigeria, all effort is geared towards promoting indigenous crop, thereby reducing total reliance on imported foods. In developing countries, where malnutrition remains a major health problem in infants and pre-school children, considerable efforts to improve the health and nutritional status of growing children have focused on the production of nutritious low-cost complementary foods (Osundahunsi and Aworh, 2003).

Maize (Zea mays) is a cereal crop widely cultivated in Nigeria and the tropics. It is used in the production of various food items such as “Ogi (Akamu/Agidi)” (fermen-
ted maize food), custard, “epiti,” (Barber et al., 2010) or
Elekute ogede” (a steamed maize-plantain pudding),
aadun (a roasted maize flour mixed with palm oil)
kokoro” (a deep fried maize paste seasoned with salt
and sugar) (Chikwendu, 2007) “Abari” also a steamed
maize pudding similar to moin-moin and “Ipekere or
Ipakere agbado” (a deep-fried maize paste seasoned
with pepper, salt and onion). Maize is sometimes used as
the starch source for beer (Maize encyclopedia, 2013).
According to FAO (1992), the protein content of maize
ranges from 8 to 11%, a variety of snack foods widely
consumed by Nigerians are made of low-protein cereals
with lysine and tryptophan as limiting amino acids
(Omueti et al., 1992) that are essential for human
nutrition.
Cowpeas (Vigna unguiculata) are an important source
of protein in developing countries especially in Nigeria. It
is an example of grain legume which has found utilization
in various ways in traditional and modern food processing
in the world (Odedeji and Oyeleke, 2011). In countries
such as Nigeria, and most of the sub-Saharan countries,
animal products representing high concentration and
quality of protein are either too expensive or simply
unaffordable, thus increasing the dependence on cereal
grains, roots and tuber crops (Ikeazor and Nogoddy,
1985). In Nigeria, cowpea is consumed in the form of
bean pudding, baked bean, fried bean, and
bean soup amongst others (Odedeji and Oyeleke,
2011). According to IITA (2009), cowpea grain contains about 25
% protein, and several vitamins and minerals. Most
tropical countries are faced with Protein-Energy
Malnutrition (PEM) as a result of increasing population
and enhanced dependence on cereal and tuber based
diet. It is estimated that about 800 million malnourished
people exist in some of the least developed countries,
mostly in sub-Saharan Africa (FAO, 2010; Meyers et al.,
1997). Cereal and legume are known to complement
each other when consumed together so as to provide
adequate nutrients for the improvement of the nutritional
well-being of the people (Chikwendu, 2007). Cowpea and
maize, in spite of their great nutritional and economic
importance are highly susceptible to many diseases and
pests right from growing stage up to storage (Singh et al.,
1997) and final consumption. Therefore, there is need to
reduce post-harvest losses of these valuable grains via
processing them into flour. There is limited information on
the nutrient composition of cereal flour enriched with
cowpea, hence, the need to go into such research.

MATERIALS AND METHODS

Materials

Dried yellow maize (Zea mays), cowpea (Vigna
unguiculata) and other ingredients (onion, fresh pepper,
palm oil) used for this research were sourced from Obada
market in Emure-ile, Owo Local Government Area of
Ondo State, Nigeria. While the AAS (Model 210 VGP
Buck Atomic Absorption Spectrophotometre) and flame
photometre used for mineral determination were from the
Chemistry Laboratory of Science Laboratory Technology
department, Hot air oven (Model Memmert 854,
Gallenkamp, UK), water bath (Model HH - 6), Kenwood
mixer (Model A 907 D, Kenwood Ltd, England) and other
equipment used for this research were from the Food
Chemistry Laboratory of the Department of Food Science
and Technology, Rufus Giwa Polytechnic, Owo, Ondo
State. All chemicals used for the analyses were of
analytical grade.

Methods

Production of maize flour

Maize flour was produced following the procedure
adopted by Barber et al., (2010) with slight modification.
The maize grains were sorted to remove extraneous
matter, then washed with potable water and boiled for 1 h
in a pressure pot. The drying was achieved with the aid of
hot air oven at 65 °C for 8 h. The dried maize was milled
into flour using attrition mill and was stored in air tight
container until needed for further analysis.

Production of Cowpea flour

Maturred and dried cowpea seeds were carefully cleaned,
sorted to remove defective ones, stones and other
extraneous matters. The cleaned seeds were soaked in
potable water for just 20 min to soften the seed coat for
easy dehulling. The dehulled cowpea was dried in hot air
oven at 65 °C for 24 h and milled into flour. The flour was
stored in air-tight polythene until needed.

Formulation

The maize:cowpea flour blends were formulated into four
ratios: 70:30; 80:20; 90:10 and 100:0 respectively, these
were kept in air tight container until needed to be used
(for analysis and thereafter production of fortified maize
snack).

Analyses

Determination of the proximate composition of the
flour blends (AOAC, 1990)

Proximate parameters including moisture, crude fat,
crude fibre, crude protein and ash contents were all
determined using the above standard method.

**Determination of carbohydrate by difference**

This is the summation of the result obtained from fat, crude fibre, ash and protein contents determination, all subtracted from 100.

\[
%\text{CHO content} = 100 - (\text{Protein} + \text{Ash} + \text{Moisture} + \text{Crude fibre} + \text{fat})
\]

**Functional properties**

**Determination of bulk density**

The bulk density (BD) was determined according to the method described by Okaka and Potter (1977). A 50 g sample was placed into a 100 ml graduated cylinder. The cylinder was tapped on the palm for 40 to 50 times and the bulk density was determined by reading the final volume. Bulk density was calculated as:

\[
\text{BD} = \frac{\text{Mass of materials}}{\text{volume of material after tapping}}
\]

**Determination of foaming capacity**

The method of Coffman and Garcia (1977) was employed in the determination of foaming capacity. Sample (1 g) was whipped with 50 ml distilled water for 5 min in a Kenwood blender at speed set at maximum and was poured into a 100 ml graduated cylinder. Total volume at time interval of 0, 5 min, 10 min up to 1 hour was noted to study the foaming capacity.

\[
%\text{Volume increase} = \frac{\text{Vol. after whipping} - \text{Vol. before whipping}}{\text{volume before whipping}} \times \frac{100}{1}
\]

**Determination of least gelation**

The modified procedure of Coffman and Garcia (1977) was used to determine gelation properties. Appropriate sample suspensions were prepared in 5 ml of distilled water each to make 2 - 20% (w/v) suspension. The test tubes containing these suspensions were heated for 1 h in boiling water (bath) followed by rapid cooling under running tap water. The test tubes were then cooled for 1 h. The least gelation concentration was determined as concentration when the sample from the inverted test tube did not fall down or slip.

\[
%\text{Least gelation} = \frac{\text{weight of sample}}{5(\text{ml})\text{of water}} \times \frac{100}{1}
\]

**Determination of water and oil absorption capacities**

Water absorption capacities (WAC) of the samples were determined by a combination of the AACC (2000) and Sathe et al. (1982) methods. Sample (1.0 g) was dispersed in 10 ml of distilled water. The content was mixed for 5 min on a magnetic stirrer or using glass rod. The mixture was centrifuged at 3,500 rpm for 30 min and the volume of the supernatant left after centrifuging was noted. Water bound was calculated from the difference in volume of the initial volume of the water used and the final volume after centrifuging. The same procedure was used for oil absorption capacity (OAC), just that oil was used in place of water.

\[
\text{WAC} = \frac{\text{Volume of water absorbed}}{\text{weight of sample}} \times \frac{100}{1}
\]

\[
\text{OAC} = \frac{\text{Volume of oil absorbed}}{\text{weight of sample}} \times \frac{100}{1}
\]

**Determination of emulsion capacity and stability**

The emulsion capacity (EC) and stability (ES) were determined by the method of Masumateu (1972). The emulsion, 20 g sample, 20 ml distilled water and 20 ml soybean oil was prepared in a calibrated centrifuge tube. The emulsion was centrifuged at 3,500 rpm for 5 min. The ratio of the height of the emulsion layer to the total height of the mixture was calculated as the emulsion activity expressed in percentage. The emulsion stability was estimated after heating the emulsion contained in a calibrated centrifuge tube at 80°C for 30 min in a water bath, cooled for 15 min under running tap water and centrifuged at 2000 rpm for 15 min. The emulsion stability expressed as a percentage was calculated as the ratio of the height of the emulsified layer to the total height of the mixture.

\[
\text{EC} = \frac{\text{height of the emulsion layer}}{\text{total height of the mixture}} \times \frac{100}{1}
\]

\[
\text{ES} = \frac{\text{height of the emulsion layer}}{\text{total height of the mixture after heating}} \times \frac{100}{1}
\]

**Determination of swelling index**

The swelling index was determined using the method of Ukpabi and Ndimele (1990). Twenty five grams of each sample were weighed into a 210 ml measuring cylinder; 150 ml of distilled water were added and allowed to stand for 4 h and the level of swelling was observed thereafter.

\[
\text{Swelling index} = \frac{\text{Volume after soaking} - \text{Volume before soaking}}{\text{weight of sample}} \times \frac{100}{1}
\]
Table 1. Proximate Composition of the Flour Blends (Dry Weight Basis).

<table>
<thead>
<tr>
<th>Samples</th>
<th>M.C(%)</th>
<th>Ash(%)</th>
<th>Fat(%)</th>
<th>Fibre(%)</th>
<th>Protein(%)</th>
<th>Carbohydrate(%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WMF</td>
<td>2.67±0.15&lt;sup&gt;c&lt;/sup&gt;</td>
<td>0.96±0.04&lt;sup&gt;d&lt;/sup&gt;</td>
<td>10.14±0.03&lt;sup&gt;b&lt;/sup&gt;</td>
<td>9.55±0.03&lt;sup&gt;d&lt;/sup&gt;</td>
<td>7.26±0.28&lt;sup&gt;d&lt;/sup&gt;</td>
<td>69.42±0.42&lt;sup&gt;a&lt;/sup&gt;</td>
</tr>
<tr>
<td>MC9</td>
<td>5.05±0.05&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.55±0.02&lt;sup&gt;c&lt;/sup&gt;</td>
<td>10.50±0.07&lt;sup&gt;a&lt;/sup&gt;</td>
<td>9.88±0.20&lt;sup&gt;c&lt;/sup&gt;</td>
<td>8.85±0.04&lt;sup&gt;c&lt;/sup&gt;</td>
<td>64.15±0.31&lt;sup&gt;b&lt;/sup&gt;</td>
</tr>
<tr>
<td>MC8</td>
<td>5.07±0.03&lt;sup&gt;b&lt;/sup&gt;</td>
<td>1.65±0.02&lt;sup&gt;c&lt;/sup&gt;</td>
<td>11.95±0.03&lt;sup&gt;a&lt;/sup&gt;</td>
<td>10.98±0.07&lt;sup&gt;a&lt;/sup&gt;</td>
<td>10.08±0.34&lt;sup&gt;b&lt;/sup&gt;</td>
<td>60.27±0.39&lt;sup&gt;c&lt;/sup&gt;</td>
</tr>
<tr>
<td>MC7</td>
<td>5.77±1.23&lt;sup&gt;c&lt;/sup&gt;</td>
<td>1.93±0.02&lt;sup&gt;a&lt;/sup&gt;</td>
<td>11.96±0.03&lt;sup&gt;a&lt;/sup&gt;</td>
<td>14.43±0.25&lt;sup&gt;a&lt;/sup&gt;</td>
<td>10.52±0.03&lt;sup&gt;a&lt;/sup&gt;</td>
<td>55.39±0.16&lt;sup&gt;d&lt;/sup&gt;</td>
</tr>
</tbody>
</table>

Values with the same superscript in the column are not significantly different (p<0.05). MC, moisture content, WMF, 100% maize – control, MC9, 10% cowpea; 90% maize, MC8, 20%cwpea; 80% maize, MC7, 30% cowpea;70% maize. Letters a-d show the degree of significant difference.

**Determination of mineral content**

The mineral contents of the samples were determined after acid digestion of the ashed samples as follows: 2 ml of aqua regia (mixture of HCl and HNO<sub>3</sub> in ratio 3:1) was added to each ashed sample in a 100 ml flask and made up to the mark with distilled water. The solution was then filtered through No.4 Whatman filter paper and the clear solution was kept in a plastic bottle with lid. Calcium, Zinc and Iron were determined using Atomic Absorption Spectrophotometre while sodium and potassium were determined using flame photometre.

**Microbiological analysis**

The media used for microbiological analysis include Nutrient agar (NA), Eosine Methylene Blue (EMB) and Potato Dextrose Agar (PDA) for total viable count, coliform test, yeast and mould count respectively. These media were obtained from the Microbiology Laboratory of the Department of Food Science and Technology, Rufus Giwa Polytechnic, Owo and were prepared according to the manufacturer’s instruction. The procedure described by Collins et al. (1989) as reported by Otunola et al. (2012) was used to evaluate the microbiological characteristics of the samples. The sample (10 g) was added to 100 ml of distilled water and mixed thoroughly after which 1 ml of the mixture was serially diluted for estimating the number of microorganisms.

**Statistical analysis**

The SPSS for windows programme version 15.0 was used to analyse the results obtained: means and standard deviation of all the samples were calculated and compared. The results obtained were in triplicate and subjected to analysis of variance ANOVA and the means were separated by New Duncan Multiple Range Test (NDMRT).

**RESULTS**

**Proximate Composition of Flour Blends**

The result of the proximate composition of the flour blends showed that there were significant differences (p<0.05) among the samples examined. The values obtained for ash ranged from 0.96 to 1.93%, fat 10.14 to 11.96%, fibre 9.55 to 14.43% and protein 7.26 to 10.52% contents also increased with increase in the cowpea substitution (Table 1). The least values were obtained in each sample for the 100% maize flour followed by 10% cowpea substitution in that order and the highest value recorded for 30% cowpea substitution. The ash content of the 100% maize flour (0.96%) was significantly different from those of the flour blends (1.55%, 1.65%, 1.93%) for 10%, 20% and 30% cowpea inclusion respectively. The fat content obtained for the flour blends (10.50%, 11.95% and 11.96% for samples 10%, 20% and 30% cowpea substitution respectively) were not significantly different among the flour blends but these values were significantly different from the fat content value obtained for the 100% maize flour (Table 1). There were also significant differences (p<0.05) among the values obtained for fibre and protein contents. While moisture, ash, fat, fibre and protein contents increased with increasing cowpea substitution, there was reduction in the carbohydrate content as the cowpea inclusion increases, and these values (69.42%, 64.15%, 60.27%, 55.39% for 100% maize flour, 10%, 20% and 30% cowpea substitution respectively) were also significantly different (p<0.05) among the samples.

**Functional Properties of Maize and Cowpea Flour Blends**

The functional properties of the flour blends showed that there were significant differences in the functional properties of the samples examined. The bulk density ranged from 0.72 g/ml to 0.81 g/ml with lowest and highest values recorded for 100% maize, 10% and 30% cowpea substitution respectively (Table 2). However, there was no significant difference (p<0.05) between the bulk densities of the samples with 10% and 30% cowpea substitution respectively. The values obtained for foaming capacity ranged from 7.67 to 11.54% with the highest value recorded for the sample with 30% cowpea substitution and the lowest value recorded for 100% maize flour. This result showed that cowpea substitution increased the foaming capacity of the flour blends, but...
the reverse was the case for water absorption capacity as this decreased with increasing cowpea substitution though with slight difference. The water absorption capacity of 100% maize flour was 2.41% which was slightly higher than the values obtained for the flour blends (3.22% and 3.23%), there was no significant difference (p < 0.05) in water absorption capacity among the flour blends (Table 2). The oil absorption capacities (OAC) of the samples also differ slightly but increased with cowpea inclusion with the highest value (1.45%) recorded for the sample with 30% cowpea substitution while the lowest value (1.24%) was recorded for 100% maize flour. The least gelation of the flour blends ranged from 6.11 to 2.1%, sample with 30% cowpea substitution had the lowest value and 100% maize had highest. These results showed that the increased in cowpea addition affect the least gelation of the flour blends by increasing the thickness of the gel. The swelling index of the flour blends was between 1.03 ml/g to 0.66 ml/g with 100% maize having the highest value and sample with 30% cowpea substitution having the least; this showed that addition of cowpea significantly reduced the swelling capacity of the flour blends. Emulsion capacity and stability were greatly influenced by the addition of cowpea in the flour blends as increase in cowpea increased the emulsion capacity and stability of the flour blends respectively. The values obtained for emulsion capacity and stability ranged from 8.76 to 3.17 x 10³ cfu/ml, the least value was recorded for 2.2 to 3.17 x 10³ cfu/ml, the yeast and mould count which ranged between 2.2

### Table 2. Functional Properties of the Flour Blends.

<table>
<thead>
<tr>
<th>Samples</th>
<th>B/D (g/ml)</th>
<th>FC (%)</th>
<th>WAC (%)</th>
<th>OAC (%)</th>
<th>LG (%)</th>
<th>SWI (ml/g)</th>
<th>EC (%)</th>
<th>ES (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WMF</td>
<td>0.72±0.01</td>
<td>7.67±0.02</td>
<td>2.41±0.02</td>
<td>1.24±0.02</td>
<td>6.11±0.11</td>
<td>1.03±0.03</td>
<td>8.76±0.02</td>
<td>7.43±0.02</td>
</tr>
<tr>
<td>MC9</td>
<td>0.81±0.02</td>
<td>11.54±0.02</td>
<td>3.22±0.02</td>
<td>1.32±0.03</td>
<td>4.10±0.01</td>
<td>0.97±0.01</td>
<td>28.15±0.17</td>
<td>12.15±0.05</td>
</tr>
<tr>
<td>MC8</td>
<td>0.76±0.02</td>
<td>15.36±0.03</td>
<td>3.23±0.02</td>
<td>1.32±0.03</td>
<td>4.06±0.05</td>
<td>0.74±0.03</td>
<td>35.08±0.07</td>
<td>13.16±0.17</td>
</tr>
<tr>
<td>MC7</td>
<td>0.81±0.01</td>
<td>11.53±0.03</td>
<td>3.23±0.02</td>
<td>1.45±0.01</td>
<td>2.10±0.1</td>
<td>0.66±0.02</td>
<td>37.15±0.16</td>
<td>14.13±0.02</td>
</tr>
</tbody>
</table>

Values with the same superscript in the column are not significantly different (p < 0.05). B/D, bulk density; FC, foaming capacity; WAC, water absorption capacity; OAC, oil absorption capacity; LG, Least gelation; SWI, swelling index; EC, emulsion capacity; ES, emulsion stability. Letters a-d show the degree of significant difference.

### Table 3. Mineral Composition of the Flour Blends.

<table>
<thead>
<tr>
<th>Samples</th>
<th>K (g/kg)</th>
<th>Na (g/kg)</th>
<th>Ca (mg/kg)</th>
<th>Zn (mg/kg)</th>
<th>Iron (mg/kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WMF</td>
<td>0.43±0.01</td>
<td>0.37±0.02</td>
<td>ND</td>
<td>0.46±0.03</td>
<td>0.88±0.02</td>
</tr>
<tr>
<td>MC9</td>
<td>0.72±0.02</td>
<td>0.37±0.01</td>
<td>3.34±0.01</td>
<td>0.77±0.20</td>
<td>0.94±0.02</td>
</tr>
<tr>
<td>MC8</td>
<td>0.89±0.01</td>
<td>0.39±0.20</td>
<td>3.36±0.02</td>
<td>1.09±0.07</td>
<td>1.17±0.02</td>
</tr>
<tr>
<td>MC7</td>
<td>0.92±0.02</td>
<td>0.43±0.02</td>
<td>3.39±0.03</td>
<td>1.13±0.02</td>
<td>1.43±0.01</td>
</tr>
</tbody>
</table>

Values with the same superscript in the column are not significantly different (p < 0.05). K, potassium; Na, sodium; Ca, calcium; Zn, zinc. Letters a-d show the degree of significant difference.

### Mineral Composition of Maize and Cowpea Flour Blends

The result of the mineral composition of the flour blends revealed significant differences (p < 0.05) between the control sample (100% maize) and the flour blends. The values obtained for potassium were 4.3 g/100 g, 7.2 g/100 g, 8.9 g/100 g and 9.2 g/100 g for 100% maize, 10%, 20%, 30% cowpea substitution respectively (Table 3). These results showed that cowpea addition increased the potassium content of the sample. The same trend was observed for sodium, zinc and iron with values ranging from 3.7 to 4.3 g/100 g, 0.46 to 1.13 mg/kg, and 0.88 to 1.43 mg/kg respectively. While calcium was not detected in the control sample (100% maize), the result obtained indicated that there was no significant difference (p < 0.05) in the calcium content among the flour blends. Addition of cowpea into maize was observed to increase the mineral contents of the flour blends.

### Microbiological Qualities of the Flour Blends

The microbiological status of the flour blends showed that the total viable count ranged from 3.8 to 4.7 x 10³ cfu/ml with the highest value recorded for 100% maize flour and the lowest value recorded for 10% cowpea substitution, with exception of this sample, there was no significant difference (p < 0.05) in the total viable count of the control sample and the flour blends (Table 4). However, significant difference was observed in the result obtained for the yeast and mould count which ranged between 2.2 to 3.17 x 10³ cfu/ml, the lowest value was recorded for...
DISCUSSION

Several data trends were observed in the result of the proximate composition of the flour blends. The moisture content of the flour blends was much higher than the value obtained for 100% maize flour. The sample with 10% cowpea substitution has the least value while the sample with 30% cowpea substitution has the highest moisture content, the values were observed to increase with increase in cowpea enrichment which implies that cowpea substitution enhanced water absorption in the sample. Similar observation was made by Barber et al. (2010). Protein content increased as a result of cowpea substitution in the flour blends, the values were significantly different among the samples. The values of fat content obtained for the flour blends were not significantly different among the flour blends but these values were significantly different (p<0.05) from the fat content value obtained for the 100% maize flour. There were also significant differences among the values obtained for ash and fibre contents. While moisture, ash, fat, fibre and protein contents increased with the increase in the cowpea fortification, there was reduction in the carbohydrate content as the cowpea inclusion increases, and these values were also significantly different (p<0.05) among the samples. Similar trend was also reported by Etsey et al. (2007).

The results of the functional properties of the flour blends showed that the cowpea substitution increased the water and oil absorption capacities as well as foaming capacity, emulsion capacity and stability. This agrees with the report of Barber et al. (2010) in the study of “effect of cowpea supplementation on the physico-chemical and sensory characteristics of epiti – a steamed maize/plantain pudding. The result for water absorption capacity agrees with the report of Afoakwa (1996) who reported that, in flour, proteins are mainly responsible for water uptake and to a lesser extent starch and cellulose at room temperature; the research of Sefa-Dedeh et al. (2001) also concurred with this. The value for oil absorption capacity increased with increase in cowpea substitution but these are lower than the water absorption capacities. The values for oil absorption capacities compared to water absorption capacities suggest that the major proteins in the flours are predominantly hydrophilic (Deshpande et al., 1983). There was a significant difference in the foaming capacities. The values obtained for the least gelation, bulk density and swelling index also showed significant difference. The emulsion capacity and stability also increased with increase in cowpea substitution respectively. These showed that emulsion capacity and stability are significantly increased with increase in cowpea addition.

There was a significant difference (p<0.05) between the control sample (100% maize) and the flour blends in terms of mineral. The values obtained for potassium in the samples was promising. These results showed that cowpea addition increases the potassium content of the sample. The same trend was observed for sodium, zinc and iron respectively. While calcium was not detected in the control sample (100% maize), the result obtained indicated that there was no significant difference in the calcium content among the flour blends. Addition of cowpea into maize was observed to increase the mineral contents of the flour blends. This is in agreement with the observation made by Chikwendu (2007) in the study of the chemical composition of Akara produced from ground bean and maize blends.

The total viable count of the flour blends was found to range from 3.8 to 4.7 x 10^3 cfu/ml with the highest value recorded for 100% maize flour and the lowest value recorded for 10% cowpea substitution, with exception of this sample, there was no significant difference in the total viable count of the control sample and the flour blends. However, significant difference (p<0.05) was observed in the result obtained for the yeast and mould count which ranged between 2.2 to 3.17 x 10^3 sfu/ml, the least value was recorded for 100% Maize flour while the sample with 30% cowpea substitution has the highest. The samples showed no growth for coliform test.

Table 4. Microbiological Status of the Flour Blends.

<table>
<thead>
<tr>
<th>Samples</th>
<th>TVC (x10^3 Cfu/ml)</th>
<th>YMC (x10^3 Sfu/ml)</th>
<th>Coliform (x10^3 Cfu/ml)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WMF</td>
<td>4.7 ± 0.17a</td>
<td>2.2 ± 0.12a</td>
<td>N/A</td>
</tr>
<tr>
<td>MC9</td>
<td>3.8 ± 0.21b</td>
<td>2.6 ± 0.10b</td>
<td>N/A</td>
</tr>
<tr>
<td>MC8</td>
<td>4.5 ± 0.12c</td>
<td>2.3 ± 0.12c</td>
<td>N/A</td>
</tr>
<tr>
<td>MC7</td>
<td>4.3 ± 0.10a</td>
<td>3.17 ± 0.15b</td>
<td>N/A</td>
</tr>
</tbody>
</table>

Values with the same superscript in the column are not significantly different (p<0.05). TVC, Total viable count, YMC, Yeast and Mould count, N/A, Not Available, WMF, 100% maize, MC9, 10% cowpea substitution, MC8, 20% cowpea substitution, MC7, 30% cowpea substitution.
Conclusion

This study has shown that increasing the percentage of cowpea flour in maize and cowpea flour blends improved the protein and mineral contents of the maize; this indicates a higher nutrient value with respect to protein and mineral contents. The functional properties (water and oil absorption, foaming capacity, emulsion capacity and stability) of the maize flour were also enhanced with the addition of cowpea as the blends had higher values compared to 100% maize flour. Microbiological qualities were not significantly affected by cowpea enrichment. Conclusively, cowpea can be added to maize to produce a composite flour of high nutritive and enhanced functional properties.

CONFLICT OF INTEREST

The authors declare that they have no conflict of interest.

REFERENCES

IITA. (2009). Cowpea Crop – IITA (International Institute of Tropical Agriculture (IITA)).