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Full Length Research

# Efficacy of warm water, Sodium hypochlorite and Trisodium phosphate on *E.coli O157: H7* and Salmonella typhimurium artificially inoculated in Telapia nilotica fillets

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ABSTRACT: The Nile tilapia (*Oreochromis niloticus*) (*Tilapia nilotica*) is a native fish in Egypt and it is widely consumed as it represent a great protein source. The health of Egyptians is badly affected by some foodborne pathogens such as *E.coli O157: H7* and *Salmonella Typhimurium* which are being transmitted by eating the contaminated *Tilapia nilotica* fillets. This experiment was carried out to decontaminate tilapia fillets samples by reference strains of *E.coli O157: H7* and *Salmonella Typhimurium*. This was done by dipping of samples in warm water (45°C) for 5 minutes, trisodium phosphate 8% for 20 minutes and sodium hypochlorite 50 ppm for 20 minutes. The count of examined *E.coli O157: H7* was decreased by 98.05%, 94.5% and 70.49% for previous treatments respectively while the count of *Salmonella Typhimurium* was decreased by 82.62%, 78.12% and 32.39% for previous treatments respectively. Some fish products with inferior quality were distributed in markets so the government must apply strict legislations on producers to ensure safe fish products for people, and must improve hygienic conditions during fishing, transportation, processing and storage of fish products.

Key words: Foodborne pathogens, E.coli, Salmonella, Tilapia nilotica, warm water, decontaminators.

## INTRODUCTION

The production of *Tilapia nilotica* in Egypt represents 58.54% of total fish production by 867,557 ton (General Authority for Fish Resources Development, 2014). Although it is necessary to ensure food safety for the health of consumers and industry, *Salmonella* spp. and *Escherichia coli* commonly detected in many fishery products (EFSA, 2010). The incidence of foodborne infections caused by bacterial pathogens continues to be a problem in industrialized nations and developing countries (Fang, 2005). These infections resulted in health and economic burdens in those countries and were especially severe in the immuno-compromised, old and young people (Bailey, 1998). Most of the reported outbreaks were caused by pathogenic bacteria,

especially E.coli O157:H7 and Salmonella (Beuchat, 1996). E.coli O157:H7 is one of the popular notorious foodborne pathogens, with an infectious dose of as low as a few hundred cells (Karmali, 2004). The most common food poisoning disease and widely distributed in the world is salmonellosis (Gomez et al., 1997). Conventional culture-based methods, involving enrichment, isolation and confirmation steps have been used for over a century due to their sensitivity, low cost. ease of use and ability to monitor cell viability (Murakami, 2012). E. coli O157:H7 and Salmonella Typhimurium are commonly found in a wide variety of raw meats, dairy products, vegetables, fish and water (Lang et al., 2004; Rhee et al., 2003; Strachan et al., 2005; Vernozy-Rozand

et al., 2005; Wachtel and Charkowski, 2002). Contamination with *E. coli O157:H7* and *Salmonella* likely occurred on farms through the use of contaminated irrigation water and manure (Franz et al., 2005; Johannessen et al., 2005).

This study was carried out to estimate the effect of warm water (45 °C) for 5 minutes, trisodium phosphate 8% for 20 minutes and sodium hypochlorite 50 ppm for 20 minutes on *Tilapia* fillets experimentally contaminated by *E.coli O157: H7* and *Salmonella Typhimurium*.

# **MATERIALS AND METHODS**

#### **Strains**

Reference strains (*E.coli O157: H7* and *Salmonella Typhimurium*) were collected from AHRI (Animal Health Research Institute) – Dokki – Giza – Egypt.

#### **Decontaminators**

Warm water 45°C for 5 minutes dipping, trisodium phosphate 8% for 20 minutes dipping and Sodium hypochlorite 50 ppm for 20 minutes dipping were used as the decontaminators.

# Fish samples

Thirty kilograms (30 kg) of *Tilapia nilotica* were collected directly from markets in Cairo and Sharkia governorates and transferred immediately as possible in an ice-box to Laboratory of Animal Health Research Institute, Dokki, Giza, Egypt and immediately processed.

#### **Procedures**

A loopful from specific strain was taken from soft agar to slope agar and then incubated for 24 hours at 37°C. Then a loopful was taken into 225 ml sterile peptone water and incubated for 24 hours at 37°C. The counts of each strain in inoculated peptone (X) (*E-coli* and *Salmonella Typhimurium*) were done.

Tilapia fillets were divided into nearly 100 gm cuts. Ten fish samples were dipped in a inoculated peptone for 15 minutes at room temperature of 25°C. Then, strain samples on sterile aluminum foil were divided into 2 groups. The first group (5 samples) was considered as inoculated samples (C) where 25 gm from each sample were added to 225 ml peptone water in stomacher bag and blended in a stomacher for 2 minutes at medium speed. The second group (5 samples for each decontaminator) is experimentally decontaminated by decontaminators for specific dipping times and strained

on sterile aluminum foil. Then, 225 ml peptone water was added to each sample in stomacher bag and blend in a stomacher for 2 minutes at medium speed.

Bacteriological count and isolation of each sample were recorded to identify each strain by morphological characters and biochemical. Control samples were plain samples before dipping in decontaminators.

# **Bacteriological examination**

#### Escherichia coli Isolation

One ml of food homogenate was inoculated from each dilution into 5 tubes of 9 ml Lauryl sulphate tryptose broth (LST). LST tubes were incubated for 48 hours at 35°C and the tubes were examined for gas. From each positive tube, a loopful of suspension was transmitted to broth of Brilliant green lactose bile and incubated for 48 hours at 35°C and then tested for gas production for Coliforn detection. From positive LST, a loopful was transferred from each positive LST to EC tube (E.coli broth). E.coli broth tubes were incubated at for 48 hours 45.5°C and tested for gas production for fecal Colifors detection. Each positive EC broth was agitated gently and transferred a loopful of positive EC broth on L-EMB agar plate (Levine's eosin-methylene blue) and then incubated at 35°C for 18 to 24 hours. Plates were examined for suspected colonies of E. coli (violet flat colonies with a dark center with or without greenish metallic sheen) (FDA, 2011). The Biochemical tests were carried out using Indole, Methyl red, VogesProskaur and Citrate utilization tests (Mac faddin, 2000).

## Salmonella typhimurium isolation

Homogenized sample was incubated for 18 hours at 37°C. One ml of pre-enriched sample was transferred to 10 ml Rappaport-Vassiliadis (RV) and then incubated for 24 hours at 42.5°C. Loopfuls from enriched (RV) broth were separately streaked onto (XLD) Xylose Lysine Desoxycholate agar and incubated for 24 hours at 37°C. Typical colonies (red with dark center) were selected and streaked onto Triple sugar iron (TSI) agar and incubated for 24 hours at 37°C (FDA, 2011). The Biochemical tests was carried out using Indole, Methyl red, Voges Proskaur, Citrate utilization and Hydrogen sulphide production tests (Quinn et al., 2002; Mac faddin, 2000).

#### Calculation of results

Calculate count as follows:

Count = 
$$\frac{C}{v (n1 + 0.1n2)d} \times n^3$$

**Table 1.** Effect of different decontaminants on *Tilapia nilotica* fillet (log<sub>10</sub> CFU/g.) artificially inoculated with *E.coli*.

Parameters	Control	Warm water	Trisodium phosphate	Sodium. hypochlorite
Minimum	6.28	4.35	4.95	5.74
Maximum	6.48	4.95	5.30	5.95
Mean ± SE	$6.36 \pm .037^{a}$	$4.65 \pm .099^{d}$	5.1±0.066°	5.83±0.039 <sup>b</sup>
Reduction count (Reduction %)		1.71 (98.05 %)	1.26 (94.5 %)	0.53(70.49 %)

Reduction Count = Control - After treatment; Reduction % = Control - After treated / Control; a-emeans carrying different superscript letters are significant (P < 0.05).

Where: C = the sum of colonies on all plates counted, v is the volume applied to each plate, n1 = the number of plates counted at the first dilution, n2 = the number of plates counted at the second dilution, n3 = the original volume of neat suspension (ie 10 for swab, 500 or 100 for other samples) and d = the dilution from which the first count was obtained eq10<sup>-2</sup> is 0.01.

## Statistical analysis

Results of microbiological analysis were reported as mean values ± standard error of mean (S.E). Microbial counts were converted to log10 CFU/g values. Statistical analysis of Data was done by using the statistical package for social sciences (SPSS Inc., Chicago, IL, USA) software. One way analysis of variance (ANOVA) at 95% level of confidence was done to determine significant differences in the microbial counts in samples. P < 0.05 was considered as significant.

# **RESULTS AND DISCUSSION**

To effectively inhibit growth of foodborne pathogens and spoilage bacteria in foods during storage, food antimicrobials have been added to food as a hurdle microbial growth, extending shelf life of various food products and also contribute positively to the sensory attributes. Among food antimicrobials, salts of organic acids (Knock et al., 2006) and several chemical solutions (aqueous) treatments such as hypochlorite (OCI<sup>-</sup>), trisodium phosphate solution (TSP) were used to eliminate pathogens from seafood (Su and Morrissey, 2003).

Commonly used chlorine compounds are liquid chlorine solution (HOCl) and hypochlorite (OCl-). More recently, chlorine dioxide (ClO<sub>2</sub>) and electrolyzed oxidizing (EO) water have also been used for this purpose. Specifically, ClO<sub>2</sub> used as an alternative to hypochlorite and chlorine in US and Europe as it has a virucidal, bactericidal and fungicidal effect. In addition, EO water has also been shown to possess strong bactericidal activity against various foodborne pathogens (Kim et al., 2000). Rajkowski (2012) showed the decrease in count of *E.coli* and *Salmonella* in Tilapia and cat fish by heat inactivation.

Results obtained (Table 1) showed that the mean value of *E.coli* count before dipping in different decontaminants was 6.36±0.037 log CFU/g (control), while after addition of decontaminants the mean values were 4.65±0.099, 5.1± .066 and 5.83± 0.039 log CFU/g respectively for warm water (45°C) for 5 minutes, trisodium phosphate 8% for 20 minutes and sodium hypochlorite 50 ppm for 20 minutes respectively. After the addition of decontaminants, the lowest *E.coli* count was 4.35 log CFU/g obtained by dipping in warm water for 5 minutes, while the highest *E.coli* count was 5.95 log CFU/g obtained by dipping in sodium hypochlorite 50 ppm for 20 minutes.

On the other hand, the most reduction count was obtained by dipping in warm water (45°C) for 5 minutes which represent 1.71 log CFU/g (98.05% reduction) followed by trisodium phosphate 8% for 20 minutes which represent 1.26 log CFU/g (94.5% reduction) then sodium hypochlorite 50 ppm for 20 minutes which represent 0.53 log CFU/g (70.49% reduction).

These results agreed with those reported by Northcutt et al. (2005), Sexton et al. (2007), Smith et al. (2007), NACMCF (2008), FDA (2009), Huang (2009), Kathleen Alonso-Hernando et al. (2012,Karuppasamy et al. (2015), while these results were lower than those obtained by Kim et al. (2004) (2.43 log CFU/g reduction count), Lee et al. (2009) (2.22log CFU/g), Xiao et al. (2011) (3.3 log CFU/g) in case of sodium hypochlorite and Purnell et al. (2014) (3.29 log CFU/g)in case of trisodium phosphate. However, these results were greater than that obtained by Purnell et al. (2014) (0.4 log CFU/g) in case of warm water. Most comparable studies applied on other food products as there were few researchers apply their studies on fish and its products.

Fecal coliform is an important group of coliform. *E. coli* are bacteria which are present naturally in the human intestinal tract. Its isolation indicates contamination of sea food samples by feces or sewage by using contaminated water, equipment and related persons (Pelczar et al., 2005).

Salmonellae Spp. are important foodborne pathogen responsible for disease in humans. It has been the leading cause of many outbreaks and infections around the world and is considered as one of the major causes of human gastroenteritis worldwide (Rasschaert et al.,

Parameters	Control	Warm water	Trisodium phosphate	Sodium hypochlorite
Minimum	4.95	4.20	4.41	4.90
Maximum	5.48	4.65	4.76	5.23
Mean ± SE	5.21±0.103a	4.45±0.073°	4.55±0.061bc	5.04±0.062a
Reduction count (Reduction %)		0.76 (82.62%)	0.66 (78.12 %)	0.17 (32.39 %)

Table 2. Effect of different decontaminants on Tilapia nilotica fillet (log<sub>10</sub> CFU/g.) artificially inoculated with Salmonella Typhimurium.

Reduction Count = Control - After treatment; Reduction % = Control - After treated / Control; <sup>a-e</sup>means carrying different superscript letters are significant (P < 0.05).

2005). Seafood contaminated by *Salmonella* (through all the periods of production from farm to markets) is the main cause of bacterial outbreaks associated with seafood in the EU (EFSA, 2010), in the US (CSPI, 2009) and in other countries worldwide.

From the results illustrated in Table 2, the minimum, maximum and mean values of control samples were 4.95, 5.48 and 5.21 $\pm$ 0.103 log CFU/g respectively. While by dipping in warm water for 5 minutes, these values were 4.20, 4.65, 4.45 $\pm$ 0.073 log CFU/g. These data also recorded the minimum, maximum and mean values of Salmonella Typhimurium after dipping in trisodium phosphate 8% for 20 minutes (4.41, 4.76, 4.55  $\pm$  0.061 log CFU/g). By using sodium hypochlorite 50 ppm for 20 minutes, the values of minimum, maximum and mean of Salmonella Typhimurium count were 4.90, 5.23 and 5.04 $\pm$ 0.062 log CFU/g respectively.

The most reduction count was obtained by dipping fish samples in warm water for 5 minutes was 0.76 log CFU/g (82.62%), then 0.66 log CFU/g (78.12%) for trisodium phosphate 8% for 20 minutes, but the lowest reduction count was 0.17 log CFU/g (32.39%) for samples treated with sodium hypochlorite 50 ppm for 20 minutes. These results were in agreement with those obtained by Yuk et al. (2008), Juneja et al. (2010), Wengian et al. (2012) and Karuppasamy et al. (2015), while these results were lower than the one obtained by Purnell et al. (2014) (2.1 and 3.29 log CFU/g reduction count) in case of using phosphate sodium hypochlorite and trisodium respectively. However, these results were higher than the one obtained by Purnell et al. (2014) (0.4 log CFU/g reduction count) in case of using warm water.

# Conclusion

Overall, foodborne pathogens as *E.coli* and *Salmonella* in fish products could be greatly decreased by using some decontaminators as warm water (45°C), trisodium phosphate 8% and sodium hypochlorite 50 ppm. These food borne pathogens represent a public health hazard, so we need a strict governmental legislations on all food dealers to apply hygienic conditions and food safety programs to prevent or reduce food contamination. These results can be used to provide safe and hygienic food handling instructions to the consumer and producers.

## **CONFLICT OF INTEREST**

The authors declare that they have no conflict of interest.

#### **REFERENCES**

Alonso-Hernando, A., Alonso-Calleja, C., & Capita, R. (2013). Growth kinetic parameters of Gram-positive and Gram-negative bacteria on poultry treated with various chemical decontaminants. *Food Contamination*, 33, 429-432.

Alonso-Hernando, A., Capita, R., & Alonso-Calleja, C. (2012).Behaviour of co-inoculated pathogenic and spoilage bacteria on poultry following several decontamination treatments. *International Journal of Food Microbiology*, 159, 152-159.

Bailey, J. S. (1998). Detection of *Salmonella* cells within 24–26 hours in poultry samples with the polymerase chain reaction BAX system. *J. Food Prot.*, 61, 792-795.

Beuchat, L. R. (1996). Pathogenic microorganisms associated with fresh produce. *J. Food Prot.*, 59, 204-216.

Center for Science in the Public Interest (CSPI) (2009).

Outbreak Alert. http://cspinet.

org/new/pdf/outbreakalertreport09 pdf (Accessed 10th November, 2011).

European Food Safety Authority (EFSA) (2010). European Centre for Disease Prevention and Control. The Community Summary Report on Trends and Sources of Zoonoses, Zoonotic Agents and Food-borne Outbreaks in the European Union in 2008. *EFSA Journal*, 8(1), 1496.

Fang, T. J. (2005). Bacterial contamination of ready-to-eat foods: concern for human toxicity. Rev. Food Nutr. Toxic., 6, 143-171.

Food and Drug Administration (FDA) (2009). Chapter V. Methods to Reduce/Eliminate Pathogens from Produce and Fresh-Cut Produce, Analysis and Evaluation of Preventive Control Measures for the Control and Reduction/Elimination of Microbial Hazards on Fresh and Fresh-Cut Produce, Washington, DC. Available online at https://www.fda.gov/Food/FoodScienceResearch/ucm091363.htm (Assessed on 8th of October, 2009).

Food and Drug Administration (FDA) (2011). Bacteriological analytical manual. Available online at https://www.fda.gov/food/foodscienceresearch/laboratorymet hods/ucm2006949.htm. (Accessed 2nd March, 2012).

Franz, E., van Diepeningen, A. D., de Vos, O. J., van Bruggen, H. C. (2005). Effects of cattle feeding regimen and soil management type on the fate of Escherichia coli O157:H7 and Salmonella enterica Serovar Typhimurium in manureamended soil, and lettuce. *Appl. Environ. Microbiol.*, 71,

- 6165-6174.
- Gomez, T. M., Motarjemi, Y., Miyagawa, S., Käferstein, F. K., & Stöhr, K. (1996). Foodborne salmonellosis. *World health statistics quarterly. Rapport trimestriel de statistiques sanitaires mondiales*, 50(1-2), 81-89.
- Huang, L. (2009). Thermal inactivation of *Listeria monocytogenes* in ground beef under isothermal and dynamic temperature conditions. *J. Food Eng.*, 90, 380-387.
- Johannessen, G. S., Bengtsson, G. B., Heier, B. T., Bredholt, S., Wasteson, Y., & Rørvic, L. M. (2005). Potential uptake of Escherichia coli O157:H7 from organic manure into crisphead lettuce. Appl. Environ. Microbiol., 71, 2221–2225.
- Juneja, V. K., Hwang, C., & Friedman, M. (2010). Thermal inactivation and post thermal treatment growth during storage of multiple Salmonella serotypes in ground beef as affected by sodium lactate and oregano oil. J. Food Sci., 75, M1-M6.
- Karmali, M. A. (2004). Infection by shiga toxin-producing Escherichia coli: an overview. Appl. Biochem. Biotechnol. B Mol. Biotechnol., 26(2), 117-122.
- Karuppasamy, K., Yadav, A. S., & Saxena, G. K. (2015). Thermal inactivation of Salmonella Enteritidis on chicken skin previously exposed to acidified Sodium chlorite or tri-sodium phosphate. *Journal of food science and technology*, 52(12), 8236-8243.
- Kathleen T. R. (2012). Thermal inactivation of Escherichia coli 0157:H7 and Salmonella on Catfish and Tilapia. Food Microbiol., 30, 427-431.
- Kim, C., Hung, Y. C., & Brackett, R. E. (2000). Efficacy of electrolyzed oxidizing (EO) and chemically modified water on different types of foodborne pathogens. *International Journal* of Food Microbiology, 61, 199-207.
- Kim, M. H., Jeong, J. W., & Cho, Y. J. (2004). Cleaning and storage effect of electrolyzed water manufactured by various electrolytic diaphragm. *Korean J. Food Preserv.*, 11, 160-169.
- Knock, R. C., Seyfert, M., Hunt, M. C., Dikeman, M. E., Mancini, R. A., Unruh, J. A., Higgins, J. J., & Monderen, R. A. (2006). Effects of potassium lactate, sodium chloride, sodium tripolyphosphate, and sodium acetate on color, color stability, and oxidative properties of injection-enhanced beef rib steaks. *Meat Sci.*, 74, 312-318.
- Lang, M. M., Harris, L. J., & Beuchat, L. R. (2004). Evaluation of inoculation method and inoculum drying time for their effects on survival and efficiency of recovery of *Escherichia coli* 0157:H7, Salmonella and Listeria monocytogenes inoculated on the surface of tomatoes. J. Food Prot., 67, 732–741.
- Lee, G. Y., Jang, H. I., Hwang, I. G., & Rhee, M. S. (2009).Prevalence and classification of pathogenic Escherichia coli isolated from fresh beef, poultry, and pork in Korea. *International Journal of Food Microbiology*, 134, 196-200.
- Mac faddin, J. F. (2000). Biochemical tests for identification medical bacteria. Warery press, INC. Baltimore, Md. 21202 USA.
- Murakami, T. (2012). Filter-based pathogen enrichment technology for detection of multiple viable foodborne pathogens in one day. *J. Food Prot.* 75(9), 1603-1610.
- National Advisory Committee on Microbiological Criteria for Foods (NACMCF) (2008). Response to the questions posed by the Food and Drug Administration and the national Marine fisheries Service regarding determination of cooking parameters for safe seafood for consumers. *J. Food Prot.*, 71, 1287-1308.
- Northcutt, J. K., Smith, D. P., Musgrove, M. T., Ingram, K. D., & Hinton, A. (2005). Microbiological impact of spray washing

- broiler carcasses using different chlorine concentrations and water temperatures. *Poult. Sci.*, 84(10), 1648-1652.
- Pelczar, M. J., Chan, E. C. S., & Noel, R. K. C. (2005). *Microbiology*. (5<sup>th</sup> Ed.) Tata mc Graw Hill, New Delhi, 571.
- Purnell, G., James, C., James, S. J., Howell, M., & Corry, J. E. L. (2014). Comparison of Acidified Sodium Chlorite, Chlorine Dioxide, Peroxyacetic Acid and Tri-Sodium Phosphate Spray Washes for Decontamination of Chicken Carcasses. Food Bioprocess Technol., 7, 2093-2101
- Quinn, P. J., Markey, B. K., Carter, M. E., Donnelly, W. J., & Leonard, F. C. (2002). Veterinary Microbiology and Microbial Diseases. Great Britain by HPG, Books Ltd., Bodmin, Cornwall, UK. Pp., 114-118.
- Rajkowski, K. T. (2012). Thermal inactivation of Escherichia coli O157:H7 and *Salmonella* on catfish and tilapia. *Food Microbiol.*, 30, 427-431.
- Rasschaert, G., Houf, K., Imberechts, H., Grijspeerdt, K., De Zutter, L., & Heyndrickx, M. (2005). Comparison of five repetitive-sequence-based PCR typing methods for molecular discrimination of salmonella Enterica isolates. J. Clin. Microbiol., 43, 3615-3623.
- Rhee, M. S., Lee, S. Y., Dougherty, R. H., & Kang, D. H. (2003). Antimicrobial effects of mustard flour and acetic acid against *Escherichia coli O157:H7*, Listeria monocytogenes, and *Salmonella enterica Serovar Typhimurium. Appl. Environ. Microbiol.*, 69, 2959–2963.
- Sexton, M., Raven, G., Holds, G., Pointon, A., Kiermeier, A., & Sumner, J. (2007).Effect of acidified sodium chlorite treatment on chickenbcarcases processed in South Australia. *Int. J. Food Microbiol.*, 115(2), 252-255.
- Smith, D. P., Northcutt, J. K., Cason, J. A., Hinton, A., Buhr, R. J., & Ingram, K. D. (2007). Effect of external or internal fecal contamination on numbers of bacteria on prechilled broiler carcasses. *Poult. Sci.*, 86(6), 1241-1244.
- Strachan, N. J. C., Doyle, M. P., Kasuga, F., Rotariu, O., & Ogden, I. D. (2005). Dose response modeling of *Escherichia coli O157:H7* incorporating data from foodborne and environmental outbreaks. *Int. J. Food Microbiol.*, 103, 35-47.
- Su, Y. C., & Morrissey, M. T. (2003). Reducing levels of *Listeria monocytogenes* contamination on raw salmon with acidified sodium chlorite. *J. Food Prot.*, 66, 812–818.
- Vernozy-Rozand, C., Mazuy-Cruchaudet, C., Bavai, C., Montet, M. P., Bonin, V., Dernburg, A., & Richard, Y. (2005). Growth and survival of Escherichia coli O157:H7 during the manufacture and ripening of raw goat milk lactic cheeses. *International Journal of Food Microbiology*, 105, 83–88.
- Wachtel, M. R., & Charkowski, A. O. (2002). Cross-contamination of lettuce with *Escherichia coli O157:H7. J. Food Prot.*, 65, 465–470.
- Wenqian, Y., Réka, Á., Dongwon, L., Seung-Cheol, L., & Hyun-Gyun, Y. (2012). Influence of lactate and acetate salt adaptation on *Salmonella Typhimurium* acid and heat resistance. *Food Microbiol.*, 30, 448-452.
- Xiao, D., Ye, R., Davidson, P. M., Hayes, D. G., Golden, D. A., & Zhong, Q. (2011). Sucrose monolaurate improves the efficacy of sodium hypochlorite against *Escherichia coli* 0157:H7 on spinach. *Int. J. Food Microbiol.*, 145, 64-68.
- Yuk, H. G., Jo, S. C., Seo, H. K., Park, S. M., & Lee, S. C. (2008). Effect of storage in juice with or without pulp and/or calcium lactate on the subsequent survival of Escherichia coli O157:H7 in simulated gastric fluid. *Int. J. Food Microbiol.*, 123, 198-203.